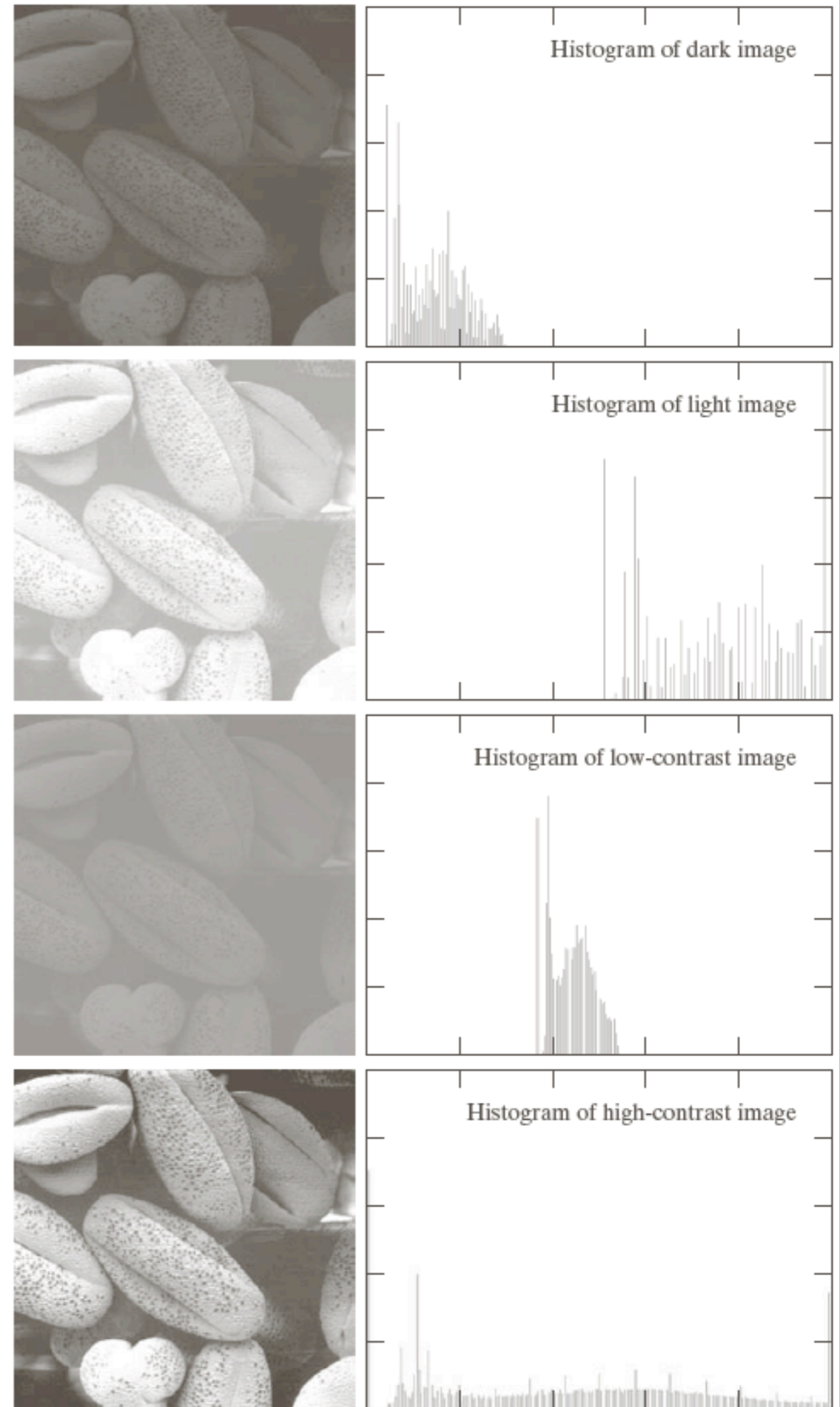


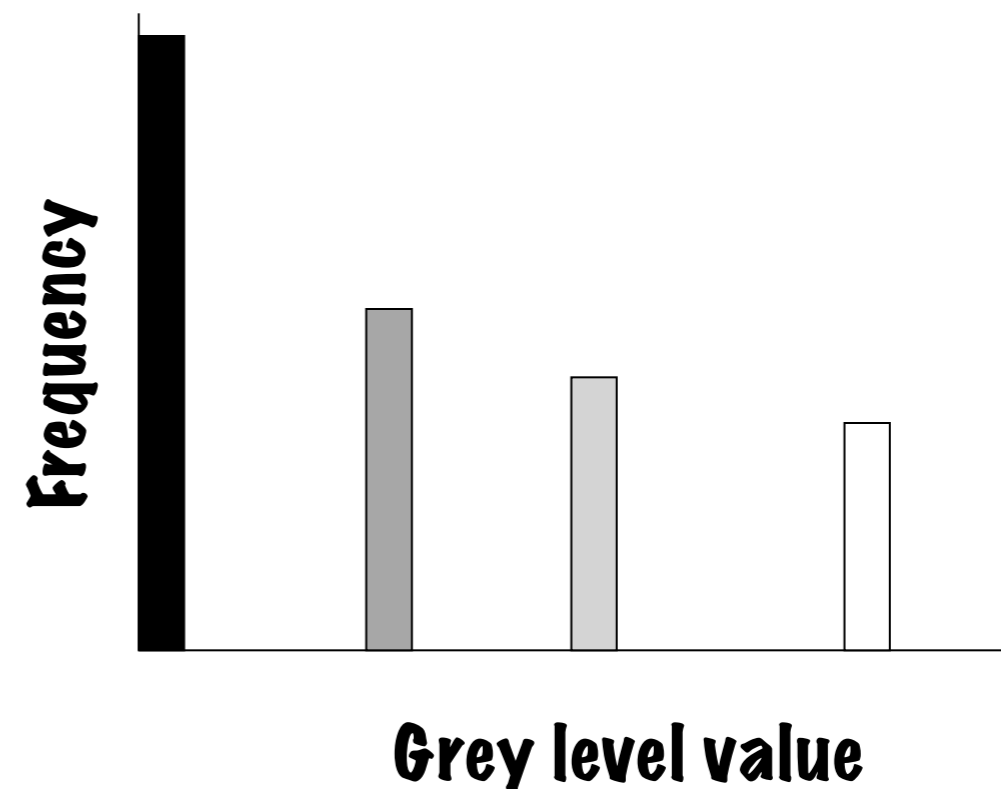
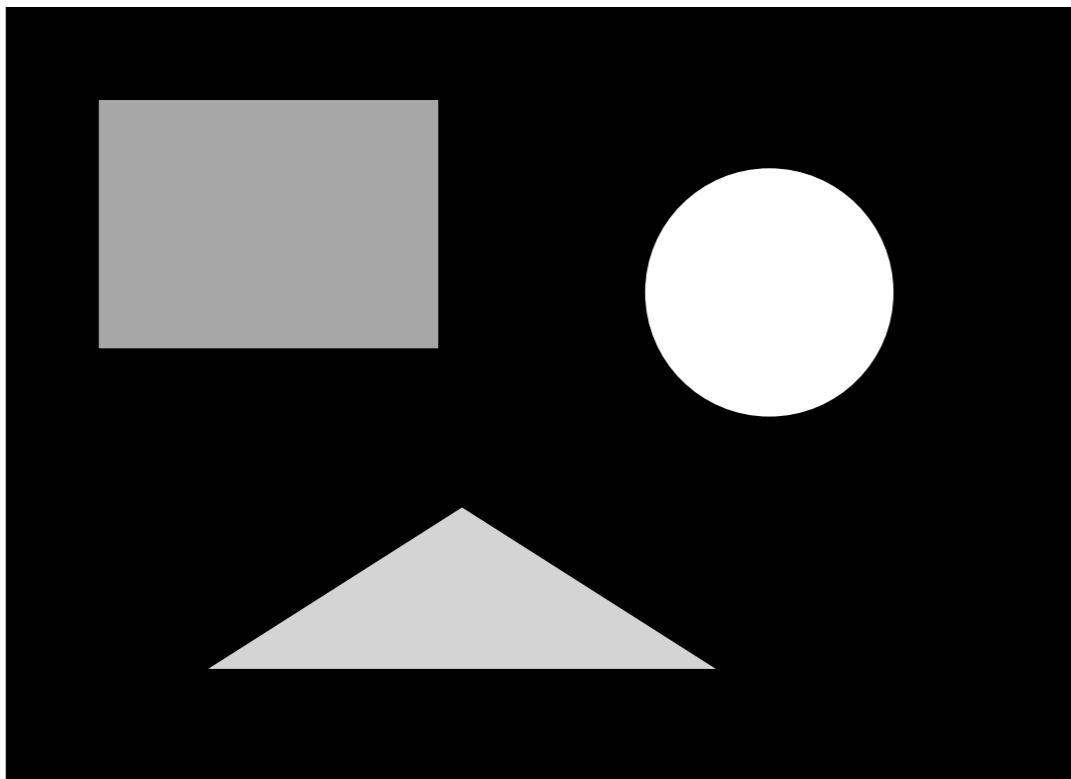
# Histograms

- $h(r_k) = n_k$ 
  - Histogram: number of times intensity level  $r_k$  appears in the image
- $p(r_k) = n_k / NM$ 
  - normalized histogram
  - also a probability of occurrence



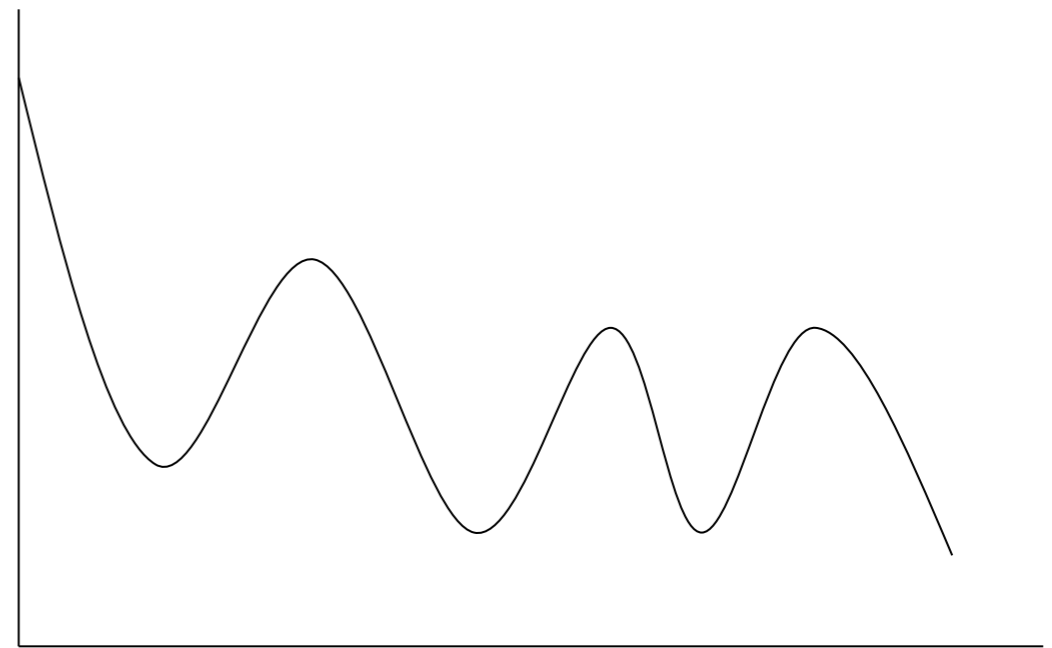
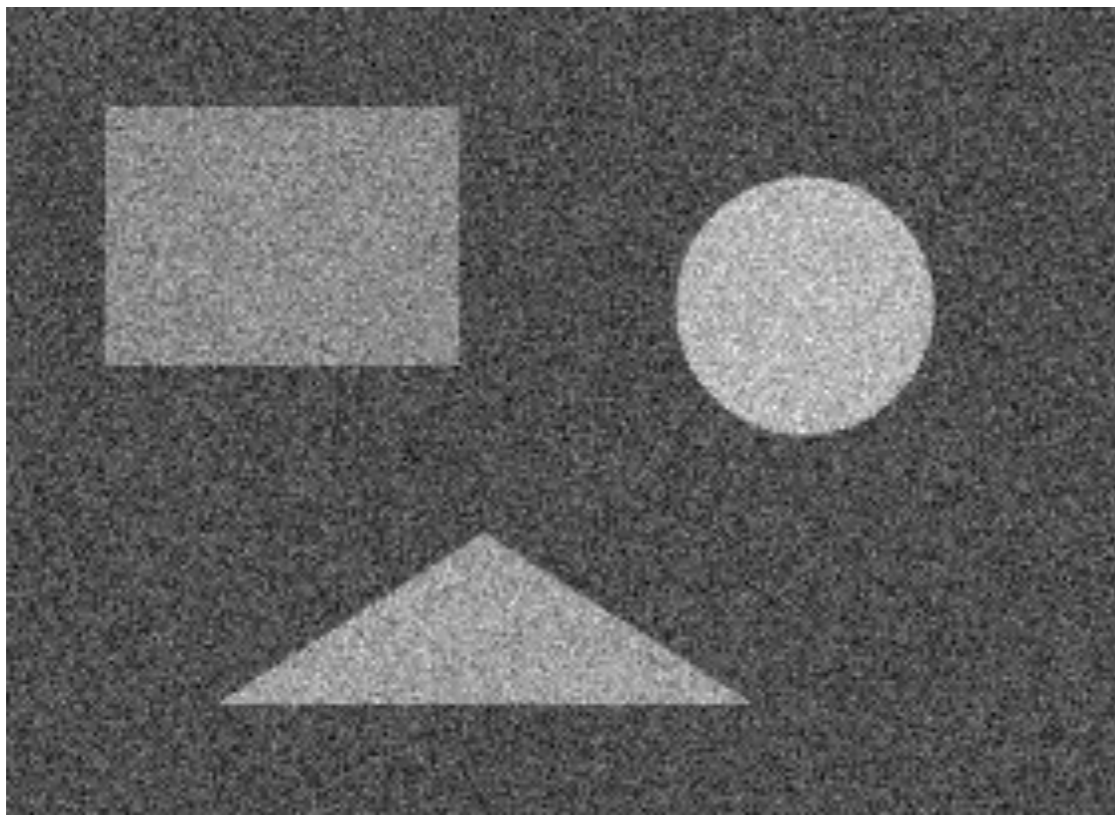
# Histogram of Image Intensities

- Create bins of intensities and count number of pixels at each level
  - Normalized (divide by total # pixels)



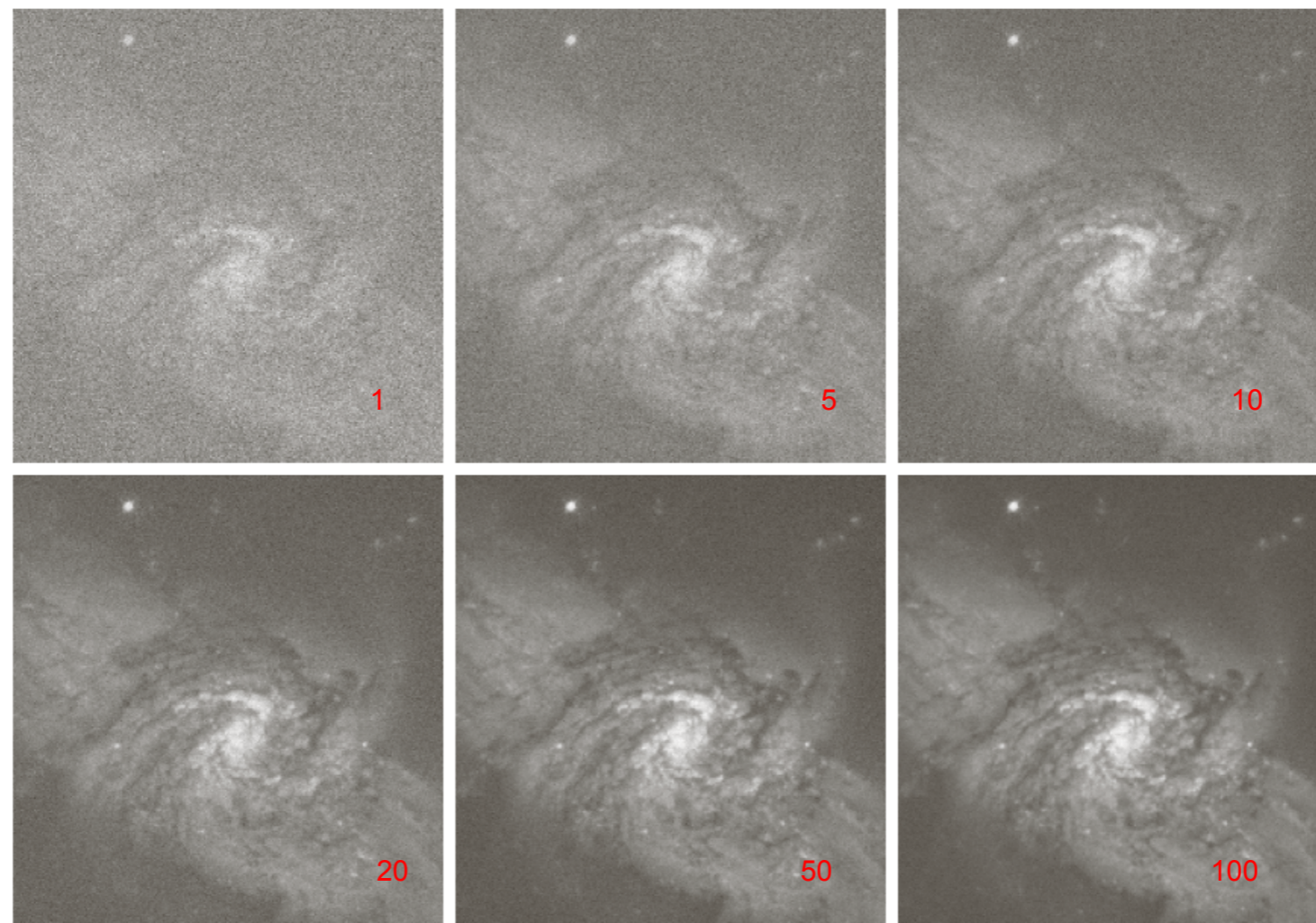
# Histograms and Noise

- What happens to the histogram if we add noise?
  - $g(x, y) = f(x, y) + n(x, y)$

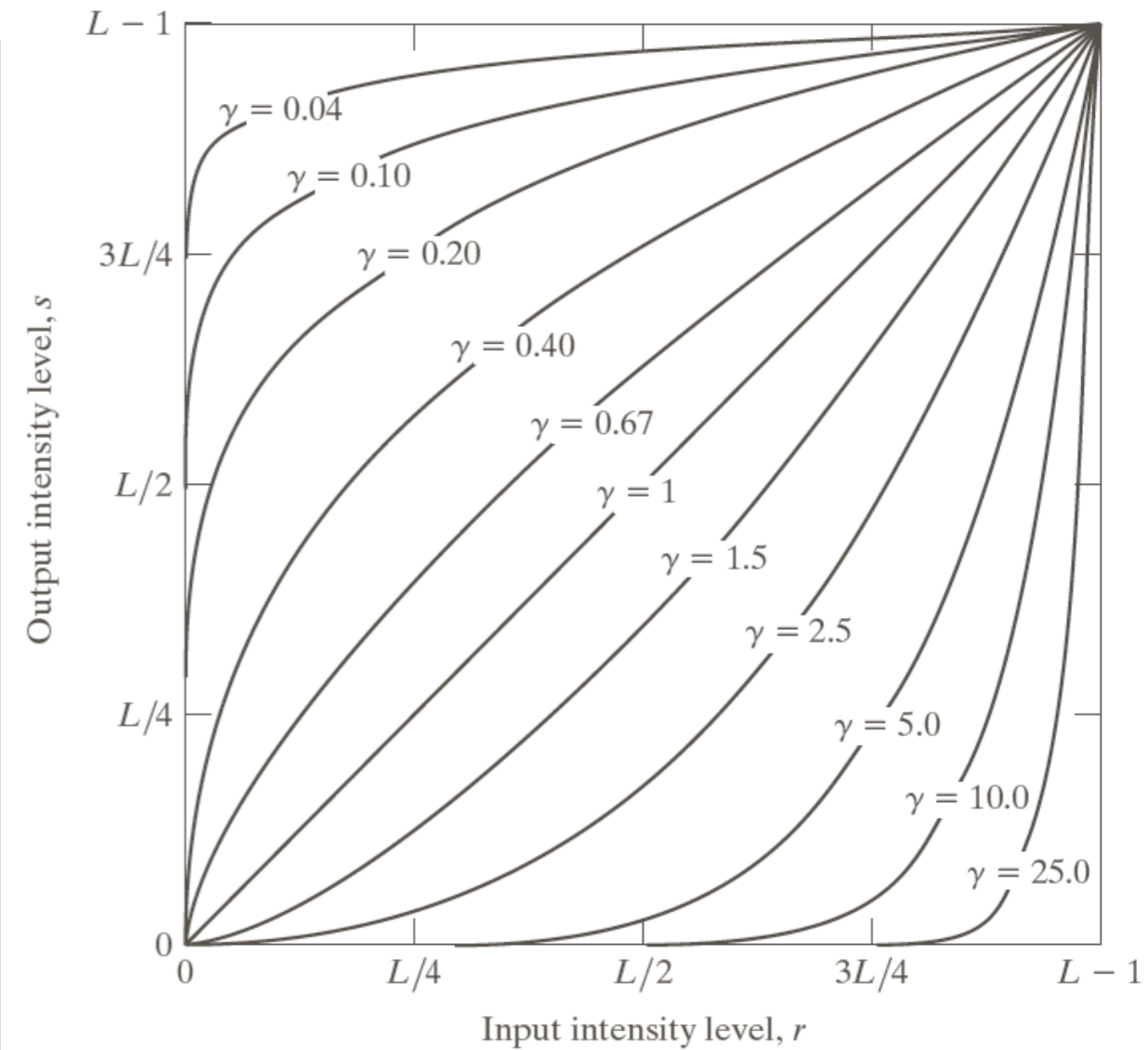
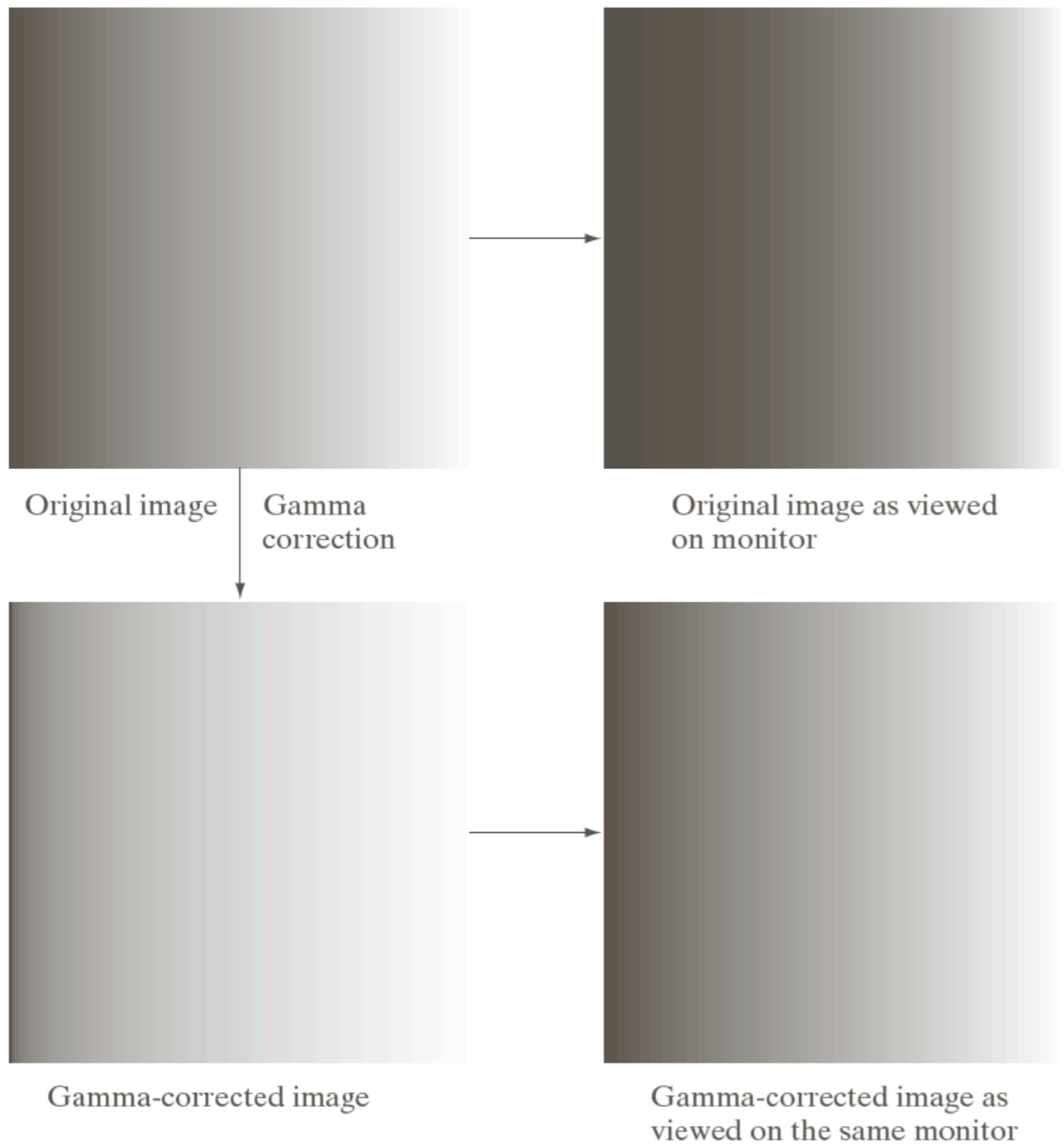


# Noise reduction with pixelwise addition

- Many noisy images of the same scene
- Averaging helps remove noise
  - Why? Under what conditions?



# Gamma correction



$$s = cr^\gamma$$

# Gamma transformations

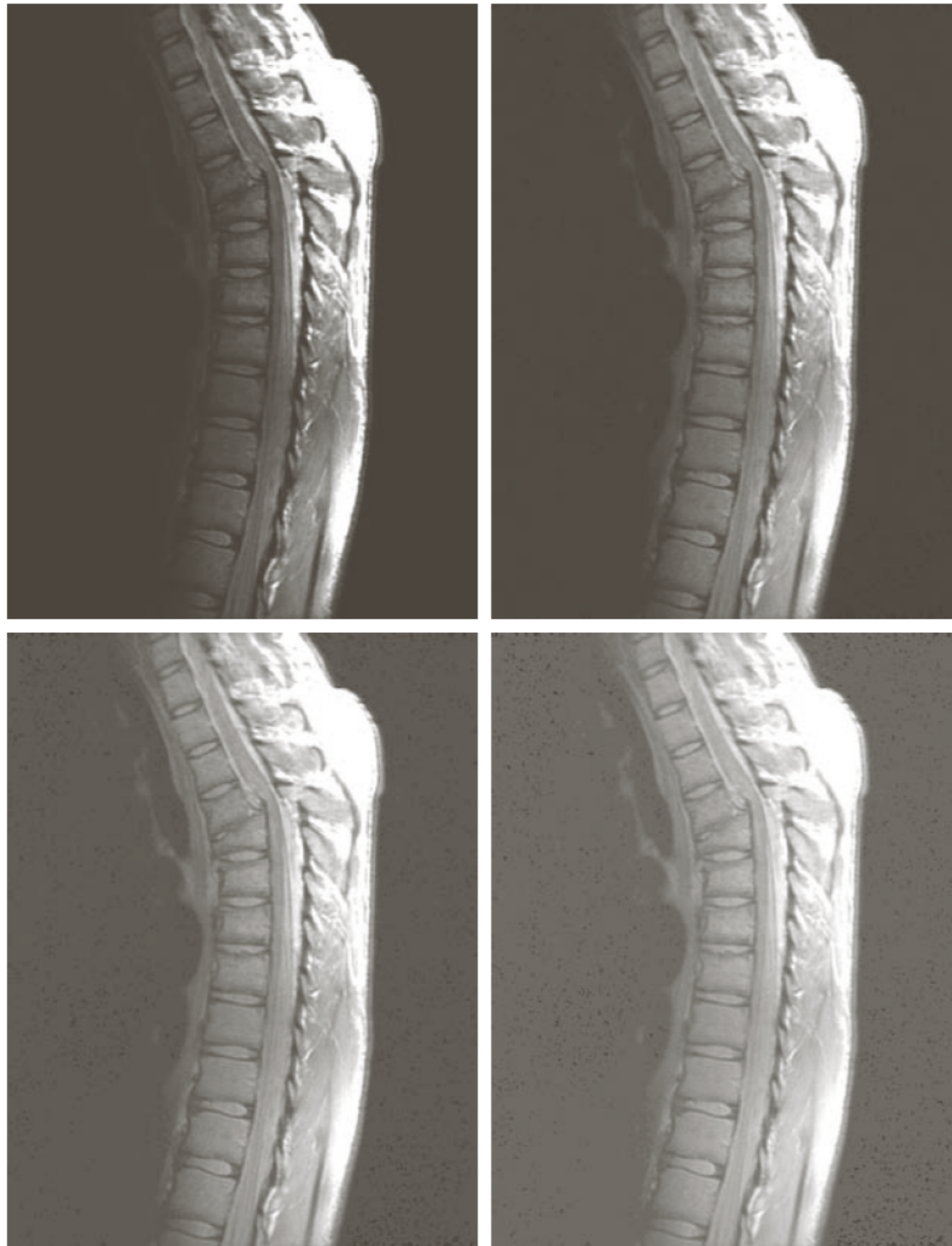


a	b
c	d

**FIGURE 3.9**

(a) Aerial image.  
(b)–(d) Results of applying the transformation in Eq. (3.2-3) with  $c = 1$  and  $\gamma = 3.0, 4.0,$  and  $5.0,$  respectively. (Original image for this example courtesy of NASA.)

# Gamma transformations



a	b
c	d

**FIGURE 3.8**

(a) Magnetic resonance image (MRI) of a fractured human spine.

(b)–(d) Results of applying the transformation in Eq. (3.2-3) with  $c = 1$  and  $\gamma = 0.6, 0.4,$  and  $0.3,$  respectively.

(Original image courtesy of Dr. David R. Pickens, Department of Radiology and Radiological Sciences, Vanderbilt University Medical Center.)

What happens to the histogram?

# Automatic histogram equalization

- Consider image intensity as a continuous valued random variable  $r$  in the interval  $[ 0 , L-1 ]$  with pdf  $p_r(r)$
- What kind of histograms do we want?
- Find an intensity transformation  $s=T(r)$  such that the pdf  $p_s(s)$  is uniform in the interval  $[ 0 , L-1 ]$
- How are the pdf's of  $r$  and  $s$  related?

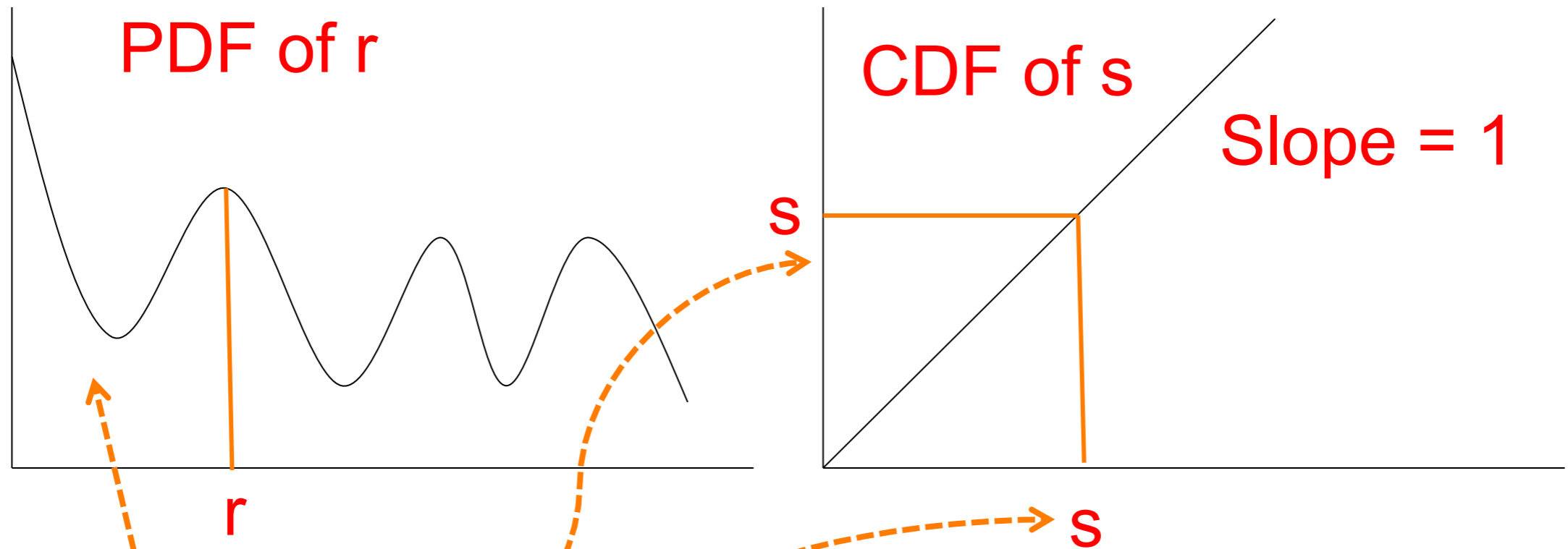
$$s = T(r) \Rightarrow p_s(s) = p_r(r) \left| \frac{dr}{ds} \right|$$



# Automatic histogram equalization

- Consider the transformation ( range [0 , 1] )

$$s = \int_0^r p_r(w) dw$$



Area given  
by integral  
above

- What is the pdf of s?
- What to do for other intensity range?

# Automatic histogram equalization

- Consider the transformation

$$s = (L - 1) \int_0^r p_r(w) dw$$

- Compute  $ds/dr$

Leibniz's rule

$$\frac{ds}{dr} = (L - 1) \frac{d}{dr} \left[ \int_0^r p_r(w) dw \right] = (L - 1) p_r(r)$$

- Verify pdf for  $s$  is uniform

$$p_s(s) = p_r(r) \left| \frac{dr}{ds} \right| = p_r(r) \left| \frac{1}{(L - 1) p_r(r)} \right| = \frac{1}{L - 1}$$

$$0 \leq s \leq L - 1$$

# (Discrete) histogram equalization

$$s_k = (L - 1) \sum_{j=0}^k p_r(r_j)$$

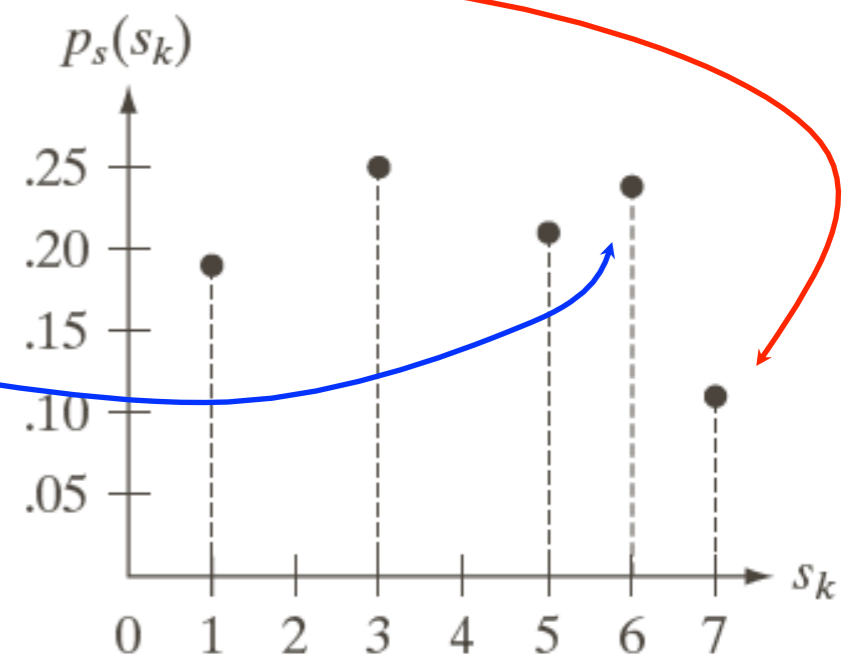
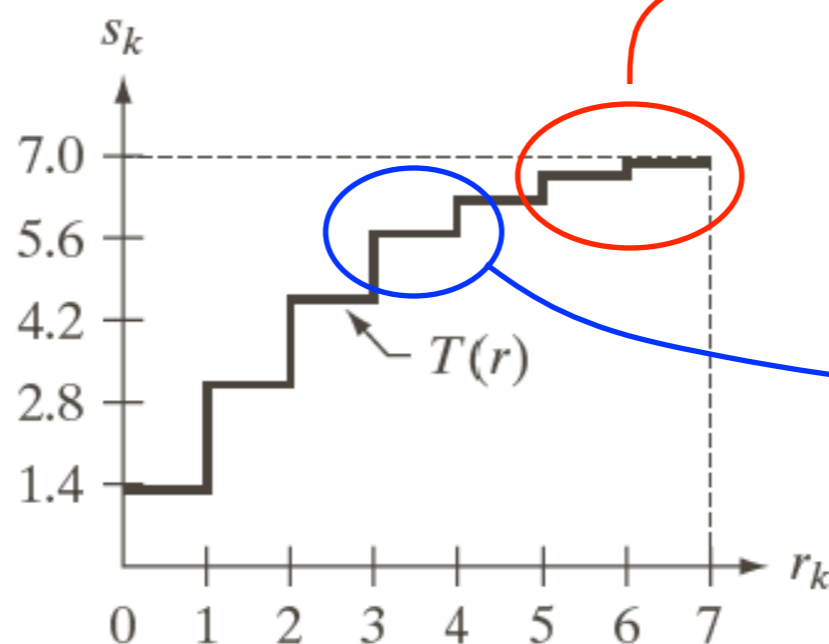
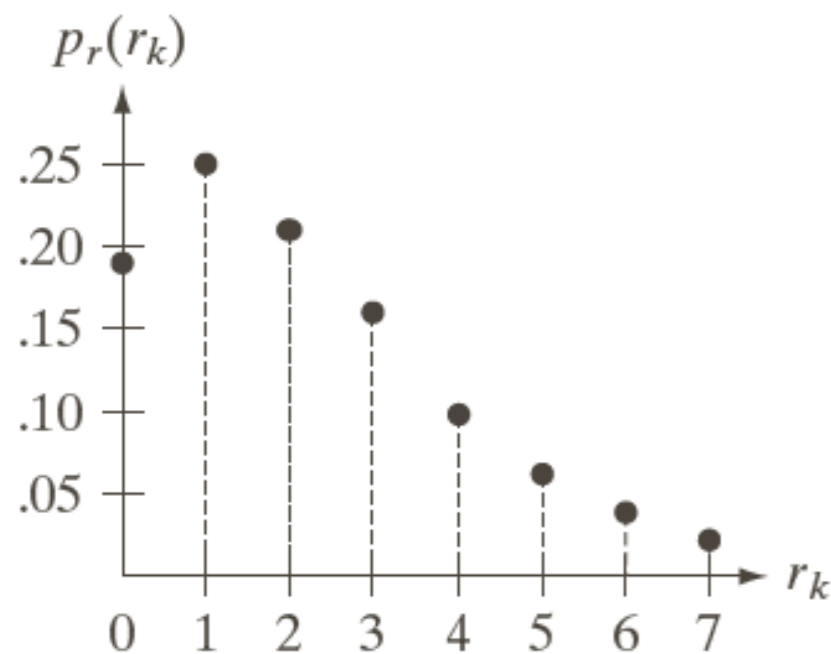
- Example with  $L=8$

$$s_0 = 7 \times 0.19 = 1.33 \rightarrow \mathbf{1}$$

$$s_1 = 7 \times (0.19 + 0.25) = 3.08 \rightarrow \mathbf{3}$$

.....

$r_k$	$n_k$	$p_r(r_k) = n_k/MN$	$S_k$
$r_0 = 0$	790	0.19	1.33
$r_1 = 1$	1023	0.25	3.08
$r_2 = 2$	850	0.21	4.55
$r_3 = 3$	656	0.16	5.67
$r_4 = 4$	329	0.08	6.23
$r_5 = 5$	245	0.06	6.65
$r_6 = 6$	122	0.03	6.86
$r_7 = 7$	81	0.02	7.00



$$S_0=1, S_1=3, S_2=5, S_3=6, S_4=6, S_5=7, S_6=7, S_7=7$$

# Histogram equalization examples



# Histogram Equalization



# Tuning Down Hist. Eq.

- Transformation is weighted combination of CDF and identity with parameter alpha

$$t(s) = (1 - \alpha)s + \alpha A(s)$$

$\alpha = 0.0$



$\alpha = 0.2$



$\alpha = 0.4$



$\alpha = 0.6$



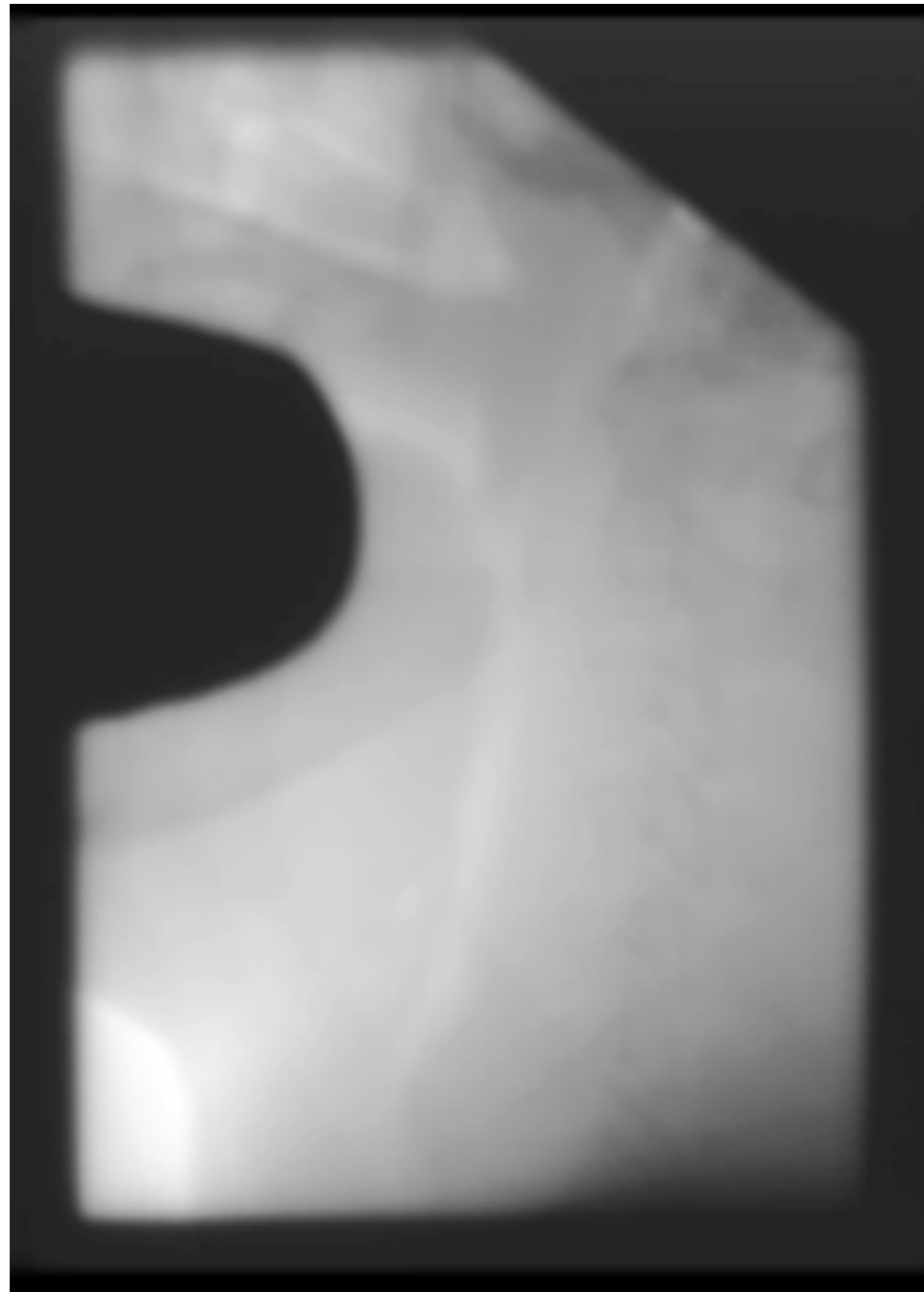
$\alpha = 0.8$



$\alpha = 1.0$



# Adaptive Histogram Equalization (AHE)



# How to do adaptive histogram equalization

- Given a local image block, either
  - Do histogram equalization for that block, OR
  - Standardize the mean and variance of each block

$$g(x, y) = 128 + 50 \frac{f(x, y) - m_B}{\sigma_B} \quad \text{for } (x, y) \in B$$

- Repeat over all blocks (non-overlapping)
- Or, Sliding window approach

$$g(x, y) = 128 + 50 \frac{f(x, y) - m_{B(x, y)}}{\sigma_{B(x, y)}}$$



# Histogram statistics

- Image  $f(x,y)$ , histogram  $p(r_i)$
- Mean intensity

$$m = \sum_{i=0}^{L-1} r_i p(r_i) = \frac{1}{MN} \sum_{x=0}^{M-1} \sum_{y=0}^{N-1} f(x,y)$$

- Second central moment (intensity variance)

$$\sigma^2 = \sum_{i=0}^{L-1} (r_i - m)^2 p(r_i) = \frac{1}{MN} \sum_{x=0}^{M-1} \sum_{y=0}^{N-1} (f(x,y) - m)^2$$

# Local histogram statistics

- $B_{xy}$  a neighborhood centered around  $(x,y)$
- We can compute the intensity histogram and its statistics (mean, variance, etc.) limited to this region

$$m_{B_{xy}} = \sum_{i=0}^{L-1} r_i p_{B_{xy}}(r_i) = \frac{1}{|B_{xy}|} \sum_{(x,y) \in B_{xy}} f(x,y)$$

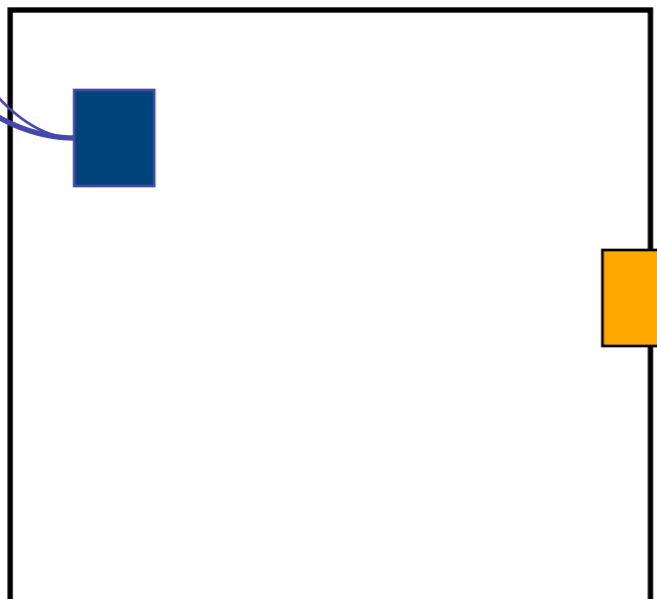
$$\sigma_{B_{xy}}^2 = \sum_{i=0}^{L-1} (r_i - m_{B_{xy}})^2 p_{B_{xy}}(r_i) = \frac{1}{|B_{xy}|} \sum_{(x,y) \in B_{xy}} (f(x,y) - m_{B_{xy}})^2$$

# Local histogram statistics

- For instance, let's define  $S_{xy}$  as a 11 x 11 neighborhood centered at  $(x,y)$ . Then

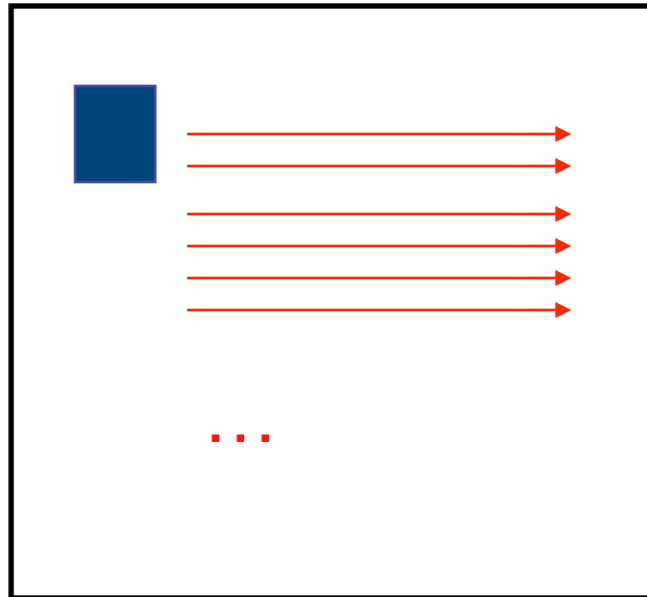
$$m_{B_{xy}} = \frac{1}{121} \sum_{u=-5}^5 \sum_{v=-5}^5 f(x+u, y+v)$$

$$\sigma_{B_{xy}}^2 = \frac{1}{121} \sum_{u=-5}^5 \sum_{v=-5}^5 \left( f(x+u, y+v) - m_{B_{xy}} \right)^2$$



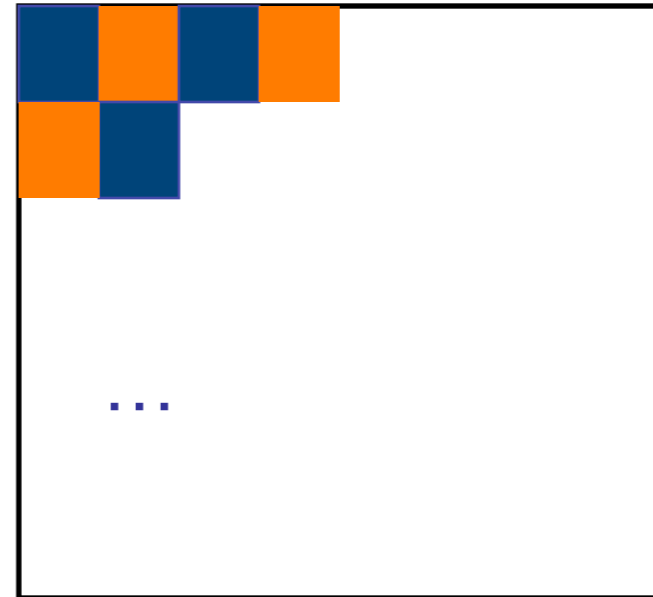
Have to take care at image boundaries

# A faster alternative



Compute statistics at every pixel

instead



Divide image into blocks. Then compute statistics

- Much more efficient
- Can create a blocky effect in the output image

# AHE Gone Bad...

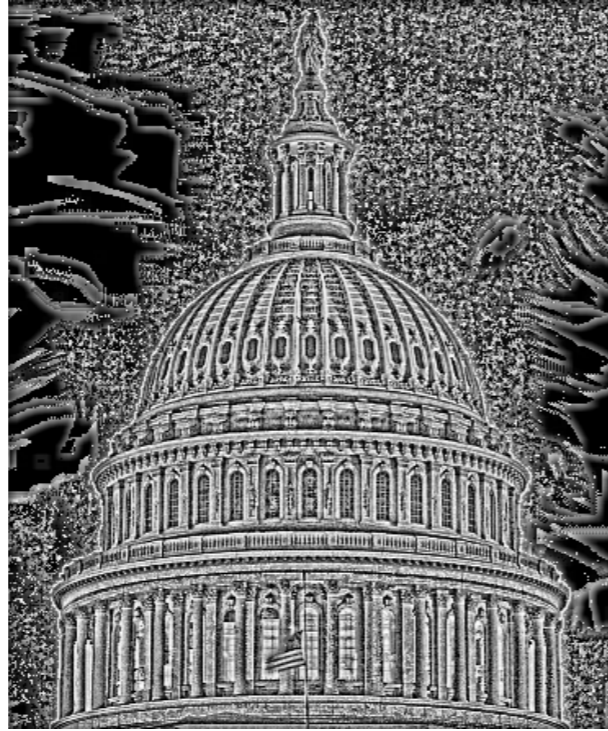


What can we do about this?

# Effect of Window Size



**Orig**



**10x10**



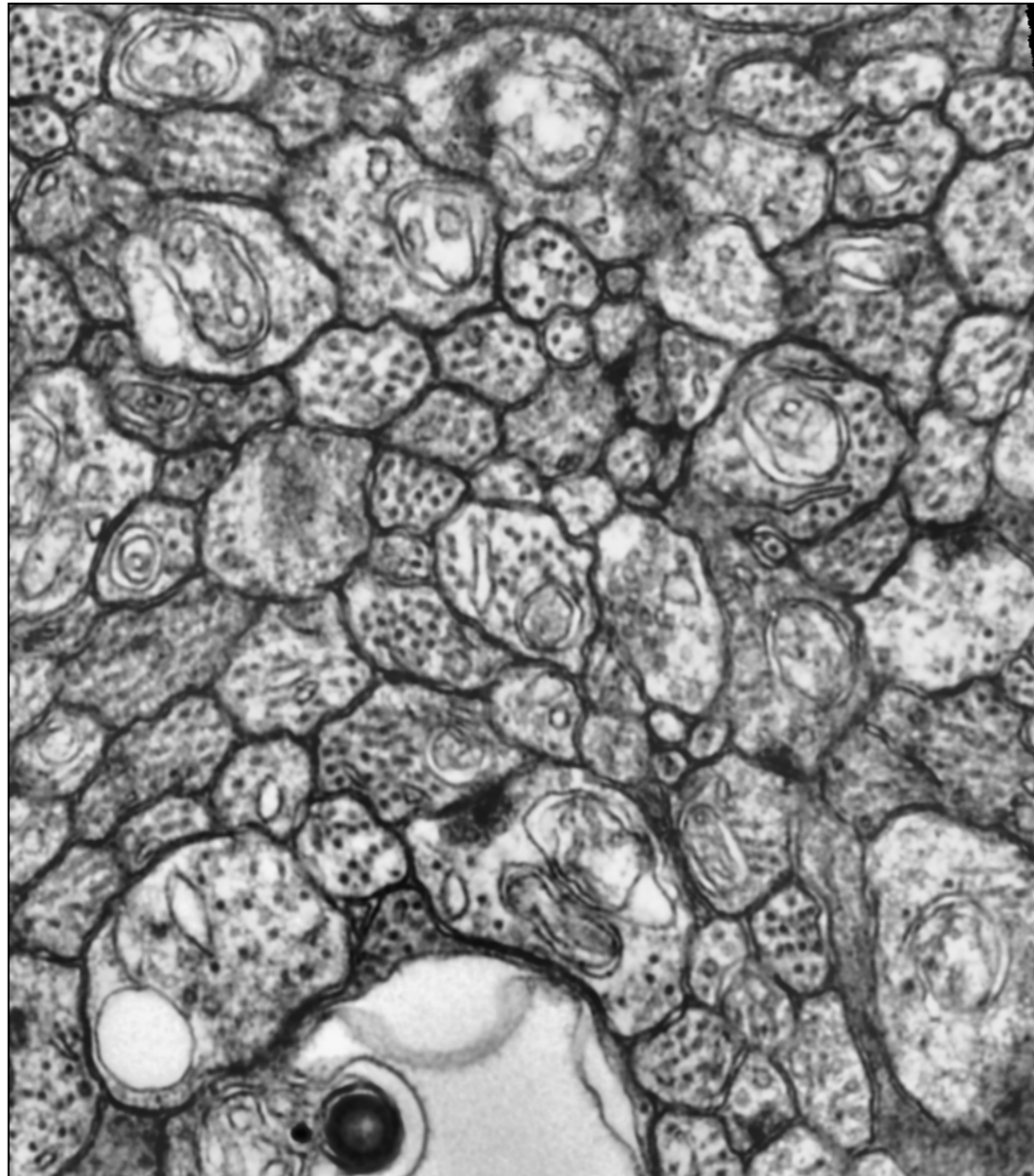
**25x25**



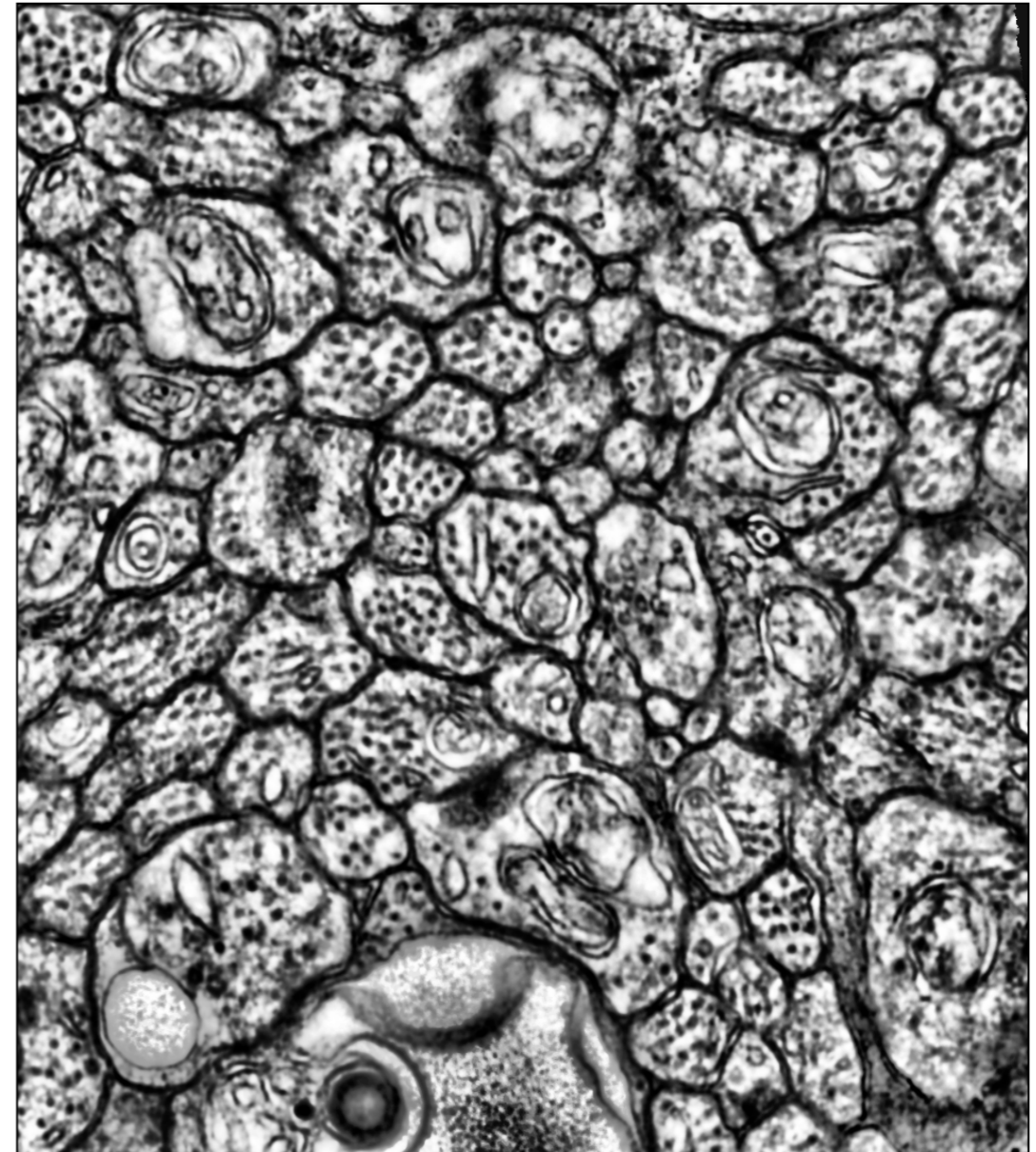
**50x50**

# AHE Application: Microscopy Imaging

**Original**



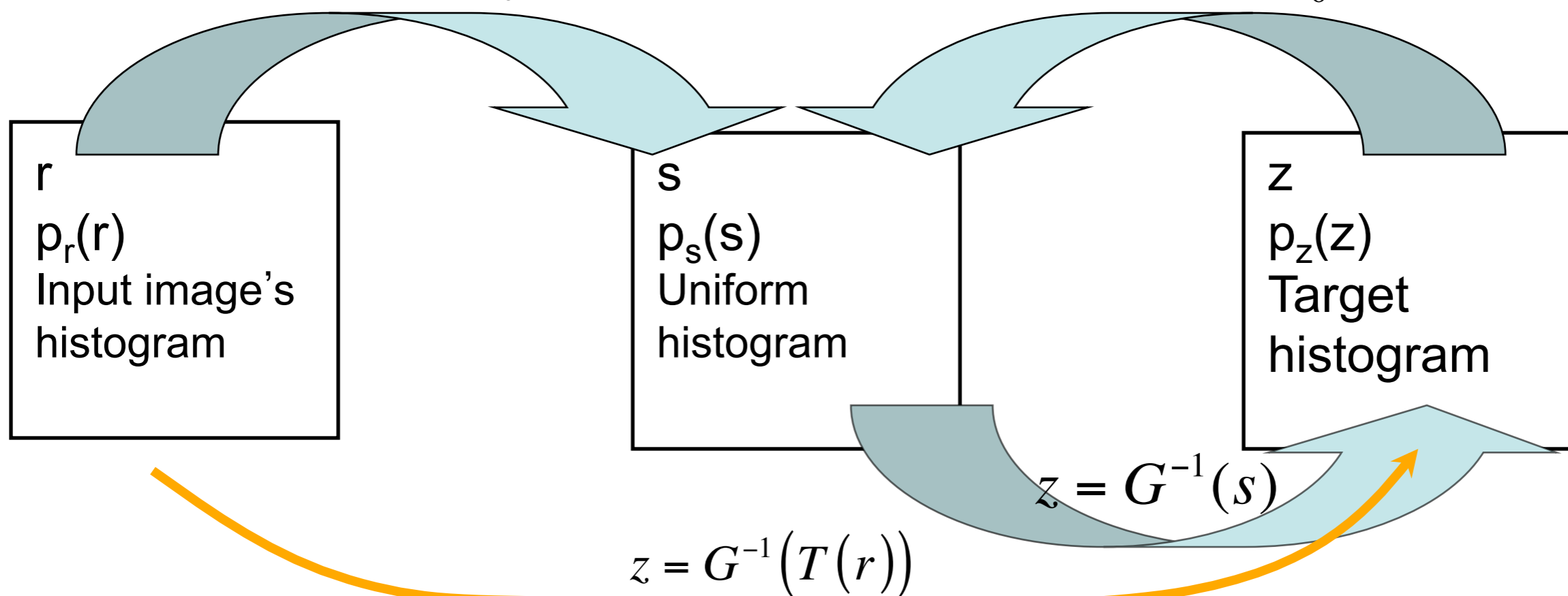
**AHE**



# Histogram matching

- Histogram equalization aims for an uniform histogram for the output image
- Sometimes we might want to specify different histograms as the target

$$s = T(r) = (L - 1) \int_0^r p_r(w) dw \quad s = G(z) = (L - 1) \int_0^z p_z(t) dt$$





# Invertible (one-to-one) transformations

- We need to be able to take the inverse of  $G(z)$

$$s = G(z) = (L - 1) \int_0^z p_z(t) dt$$

- $G(z)$  has an inverse if it is strictly monotonically increasing as a function of  $z$ 
  - This is true if  $p_z(z) > 0$  for all  $z$

# Histogram matching (discrete)

1. Compute  $G(z_q) = (L-1) \sum_{i=0}^q p_z(z_i)$

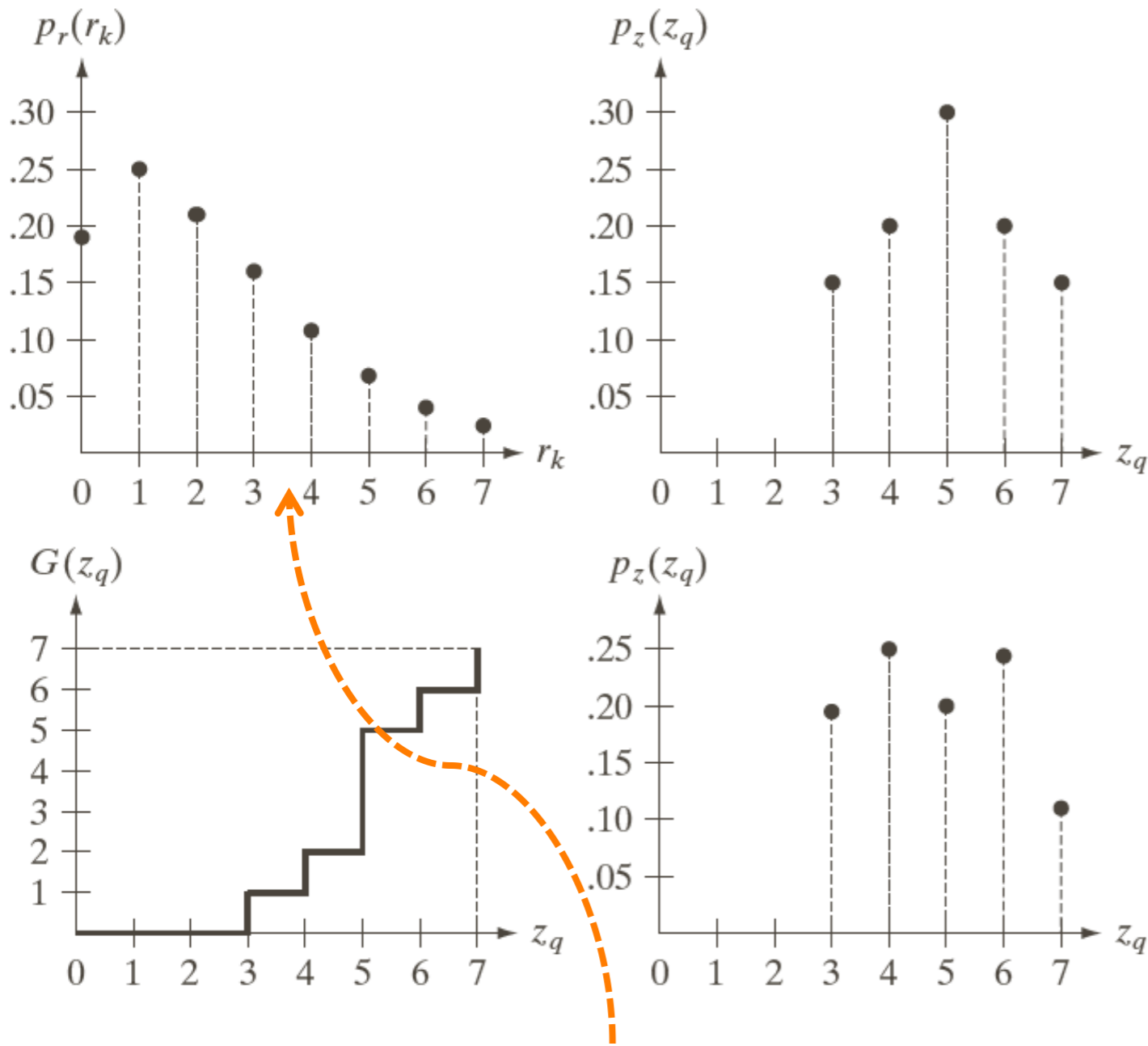
and store in a table

2. Compute  $s_k = T(r_k) = (L-1) \sum_{i=0}^k p_r(r_j)$

3. Create mapping from  $s_k$  to  $z_q$  (inverse of  $G$ ) by finding the closest value in the table stored from step 1 to all  $s_k$ .

4. Apply mapping created in step 3 to histogram equalized input image

# Example



**1**

$z_q$	Specified $p_z(z_q)$	Actual $p_z(z_k)$	G
$z_0 = 0$	0.00	0.00	0.00
$z_1 = 1$	0.00	0.00	0.00
$z_2 = 2$	0.00	0.00	0.00
$z_3 = 3$	0.15	0.19	1.05
$z_4 = 4$	0.20	0.25	2.45
$z_5 = 5$	0.30	0.21	4.55
$z_6 = 6$	0.20	0.24	5.95
$z_7 = 7$	0.15	0.11	7.00

Inverse of G

**3**

$s_k$	$\rightarrow$	$z_q$
1	$\rightarrow$	3
3	$\rightarrow$	4
5	$\rightarrow$	5
6	$\rightarrow$	6
7	$\rightarrow$	7

**2** Did this one before  
 $S_0=1$  ,  $S_1=3$  ,  $S_2=5$  ,  $S_3=6$  ,  
 $S_4=6$  ,  $S_5=7$  ,  $S_6=7$  ,  $S_7=7$

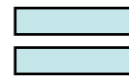
**4.**  $r_k \rightarrow s_k \rightarrow z_q$

# Colorization example



Match histograms of the gray scale input image to histograms of the Red, Green and Blue channels of a reference image individually to create 3 new images.

Use these new images as red, green blue channels to create the output color image.



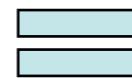
To get realistic results input and reference image contents should be similar.



# MATLAB code

- Given a gray scale image I2gray and a reference image I1color
- Uses image processing toolbox command imhistmatch
- I2red = imhistmatch(uint8(I2gray),I1color(:, :, 1));
- I2blue = imhistmatch(uint8(I2gray),I1color(:, :, 3));
- I2green = imhistmatch(uint8(I2gray),I1color(:, :, 2));
- I2colorized(:, :, 1)=I2red;
- I2colorized(:, :, 2)=I2green;
- I2colorized(:, :, 3)=I2blue;

# Colorization example

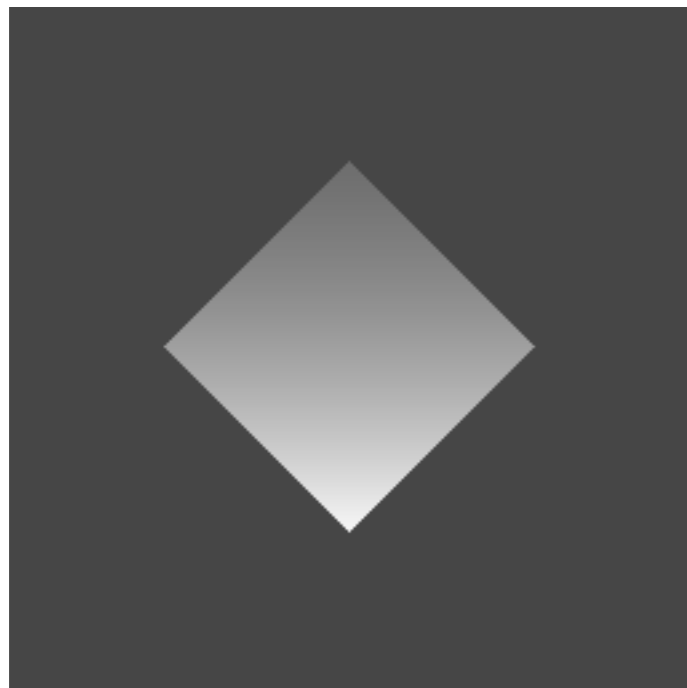


# Other uses of histograms

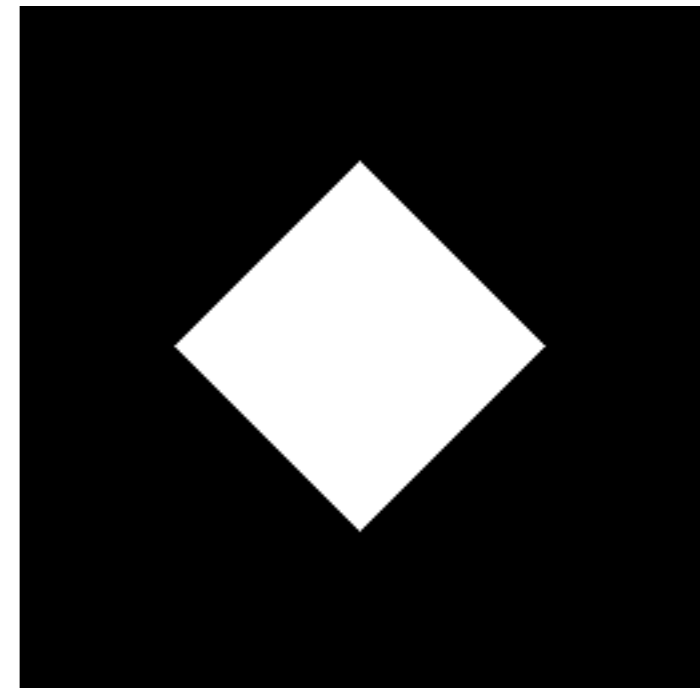
- So far we used histograms for image enhancement purposes
- Quantization
  - Reducing the number of gray levels or colors in an image for efficient storage
- Segmentation
  - Image segmentation is the process of subdividing an image into its constituent regions or objects.

# What is image segmentation?

- Image segmentation is the process of subdividing an image into its constituent regions or objects.
- Example segmentation with two regions:



Input image  
intensities 0-255



Segmentation output  
0 (background)  
1 (foreground)



# Formal definition of segmentation

- $R$ : set of all pixels in given image
- Segmentation into  $n$  regions  $R_1, R_2, \dots, R_n$
- Two important properties

– Complete

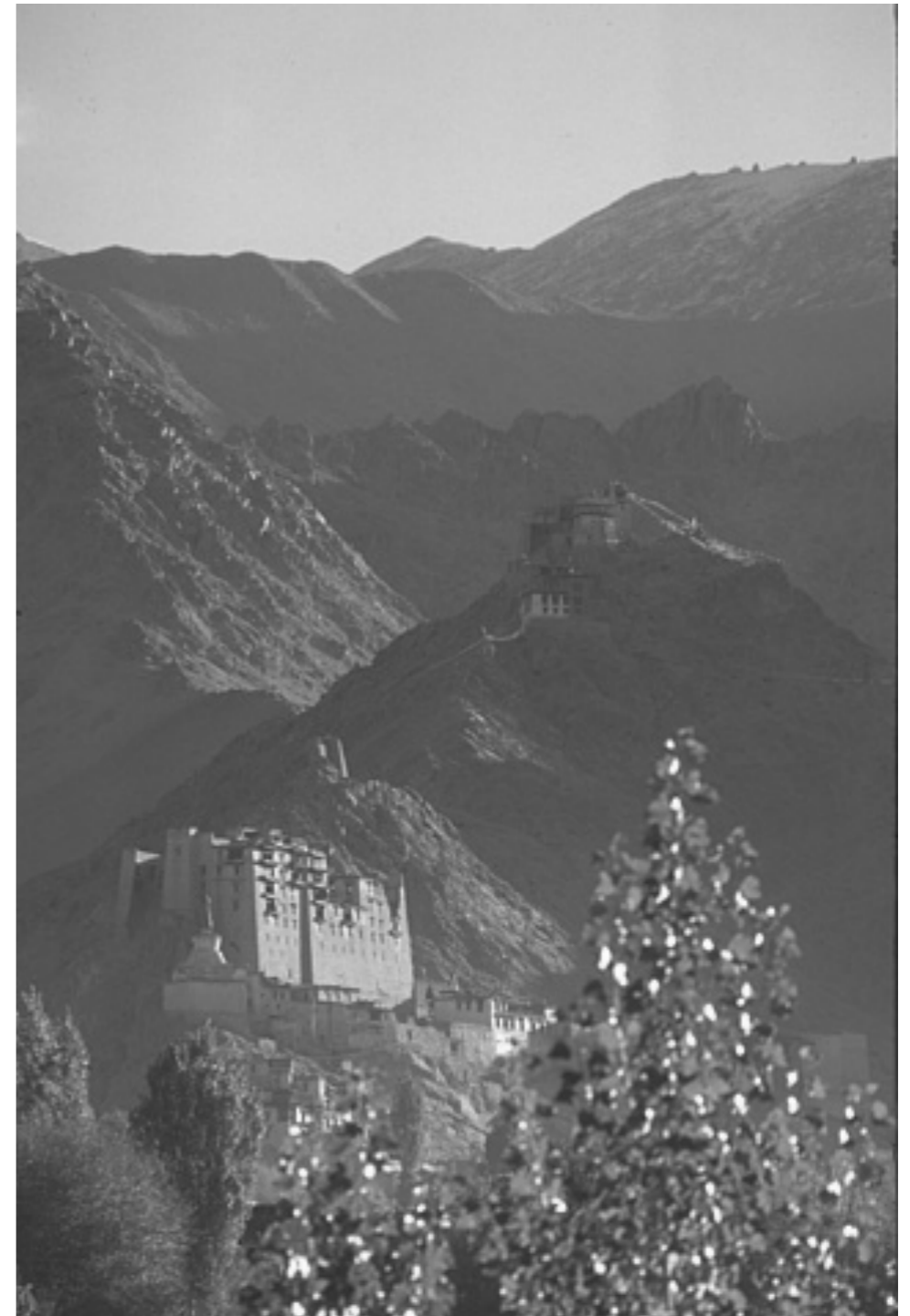
$$\bigcup_{i=1}^n R_i = R$$

– Mutually exclusive

$$R_i \cap R_j = \emptyset \text{ if } i \neq j$$

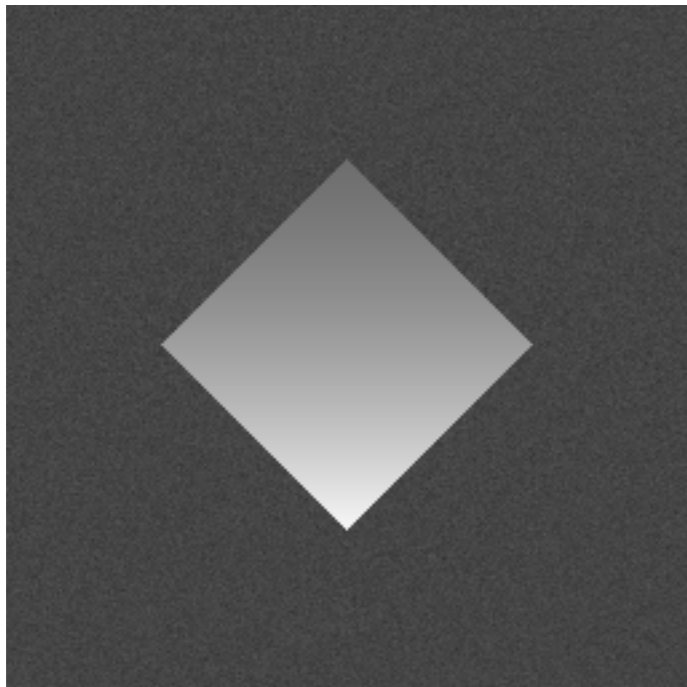
# Two general approaches to segmentation

- Discontinuity based
  - Partition an image based on abrupt changes of intensity. Find pixels which correspond to these abrupt changes, these will be the boundaries between regions.
  - Edge detection is an example of this type of approach to segmentation
- Similarity based
  - Group pixels into regions of similar intensity
  - Thresholding is an example of this type of approach to segmentation

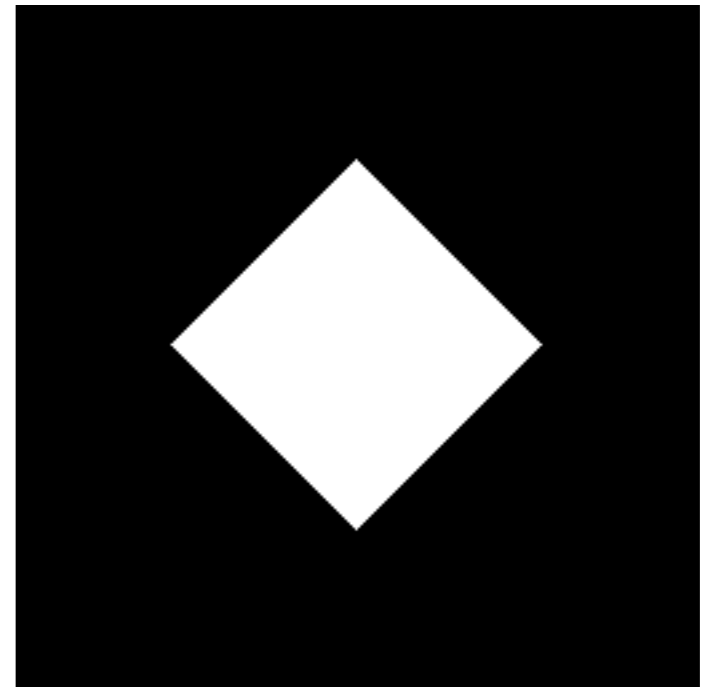


# Global thresholding

$$g(x, y) = \begin{cases} 1 & \text{if } f(x, y) > T \\ 0 & \text{if } f(x, y) \leq T \end{cases}$$



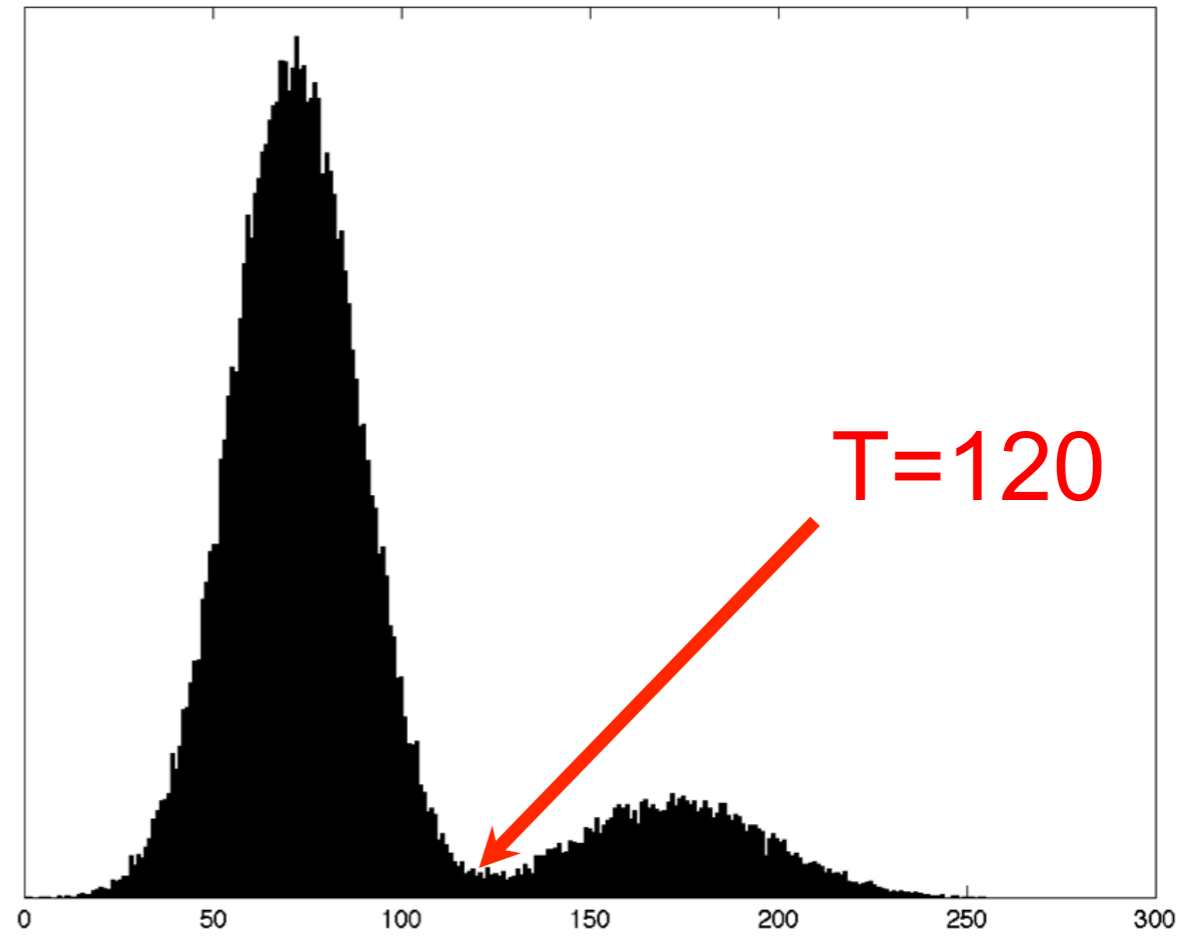
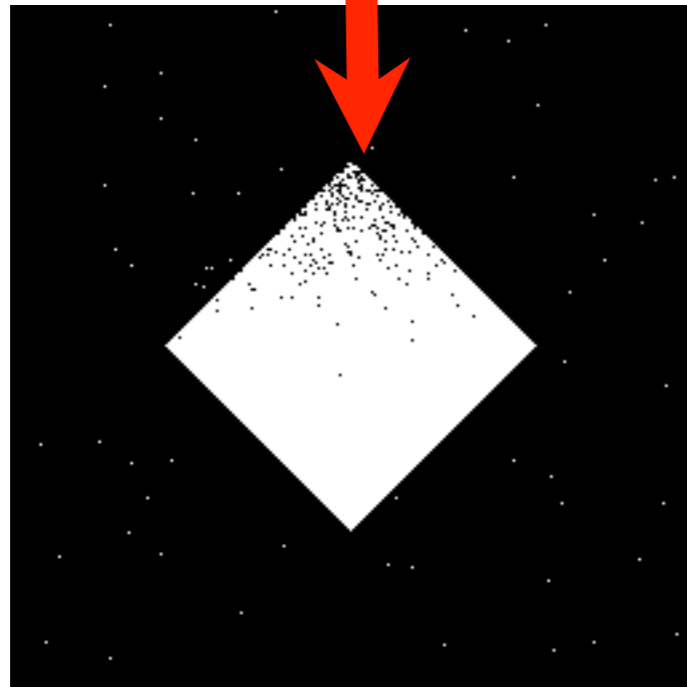
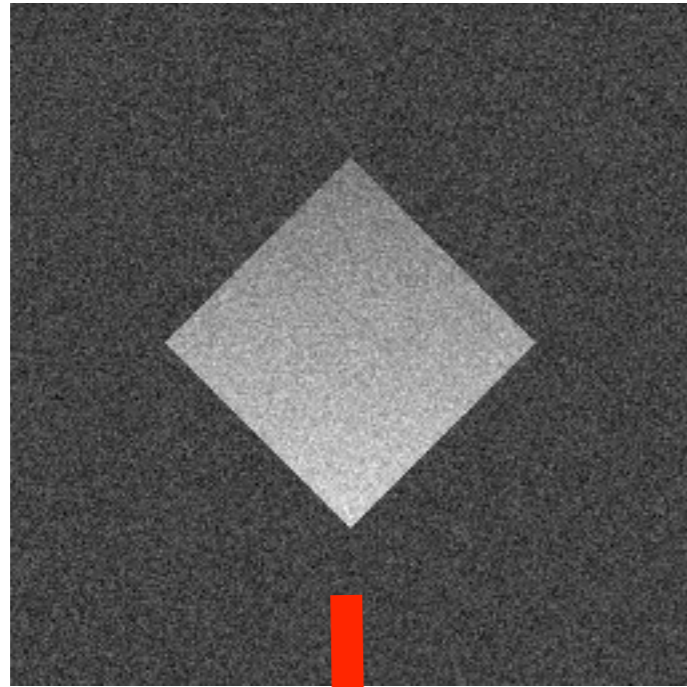
Input image  $f(x,y)$   
intensities 0-255



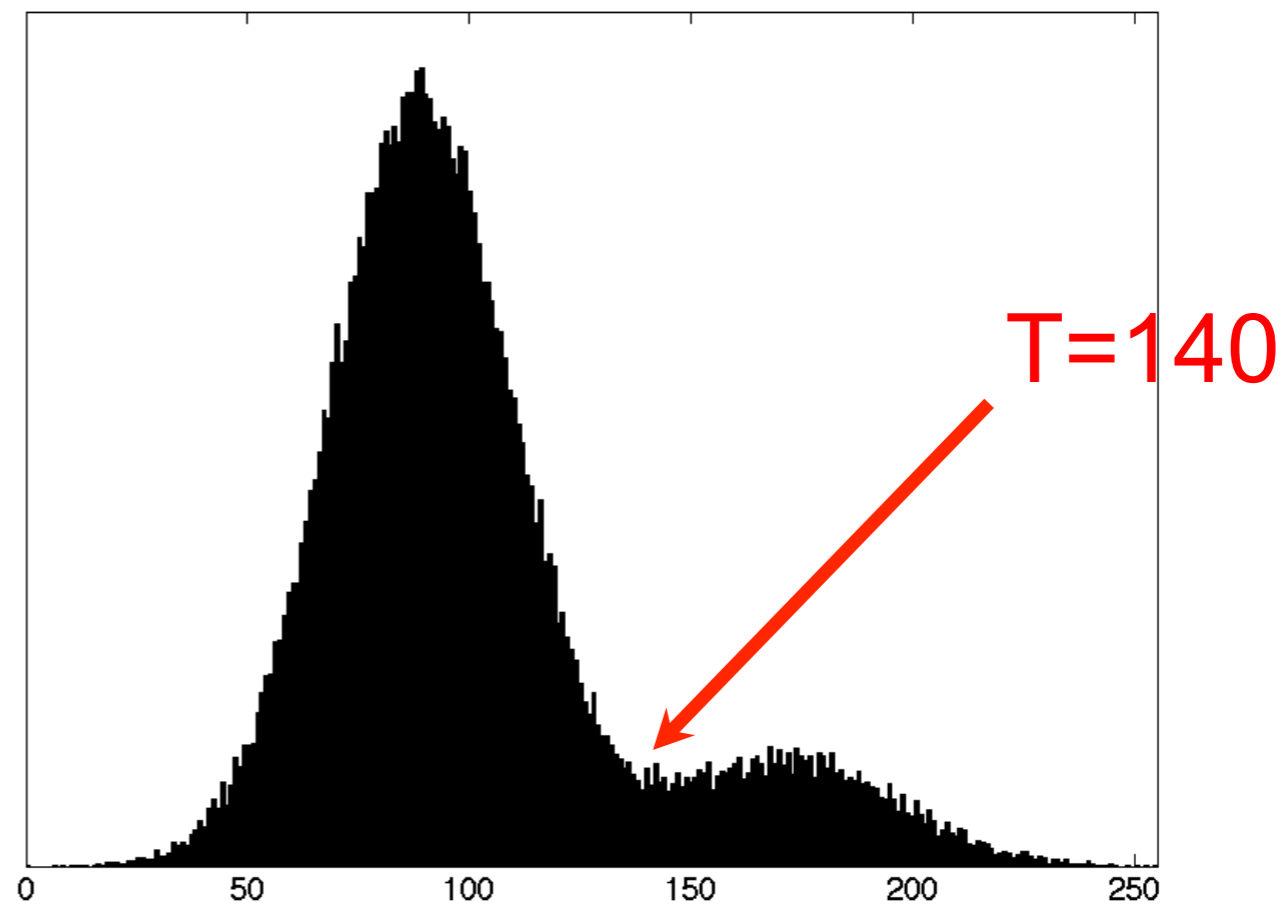
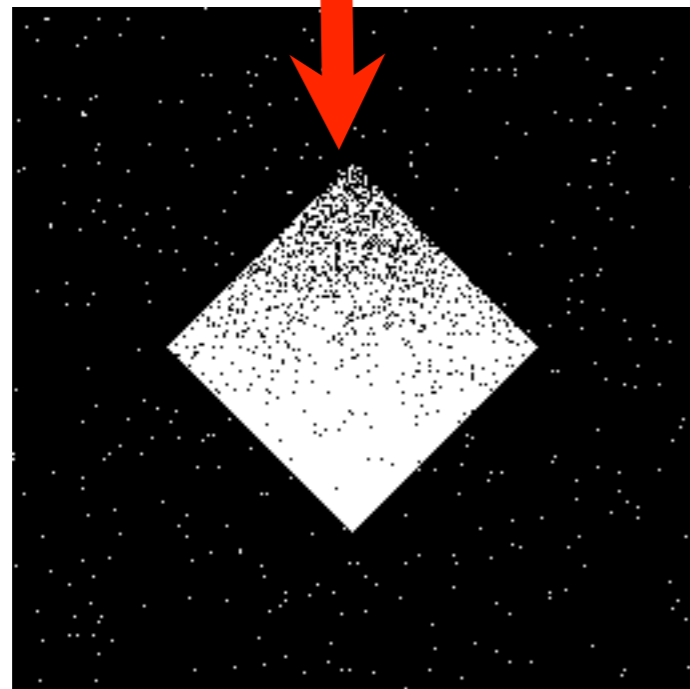
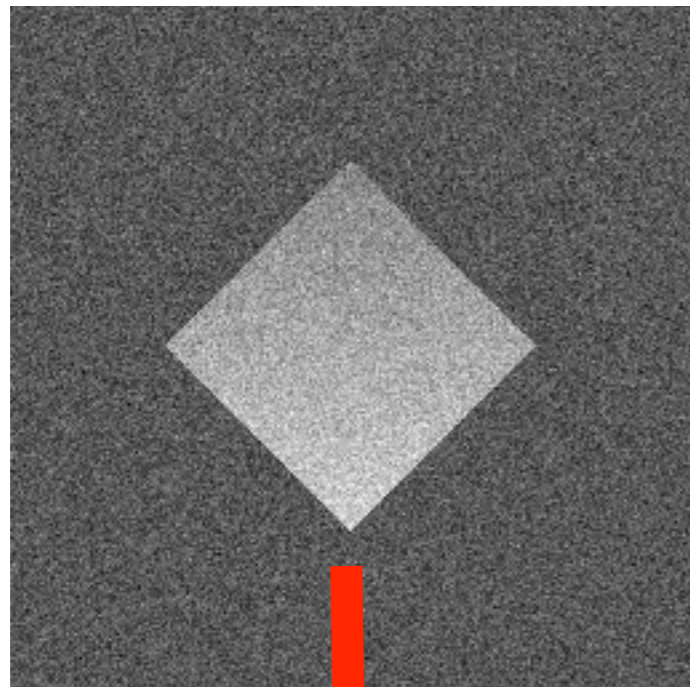
Segmentation output  $g(x,y)$   
0 (background)  
1 (foreground)

- How can we choose  $T$ ?
  - Trial and error
  - Use the histogram of  $f(x,y)$

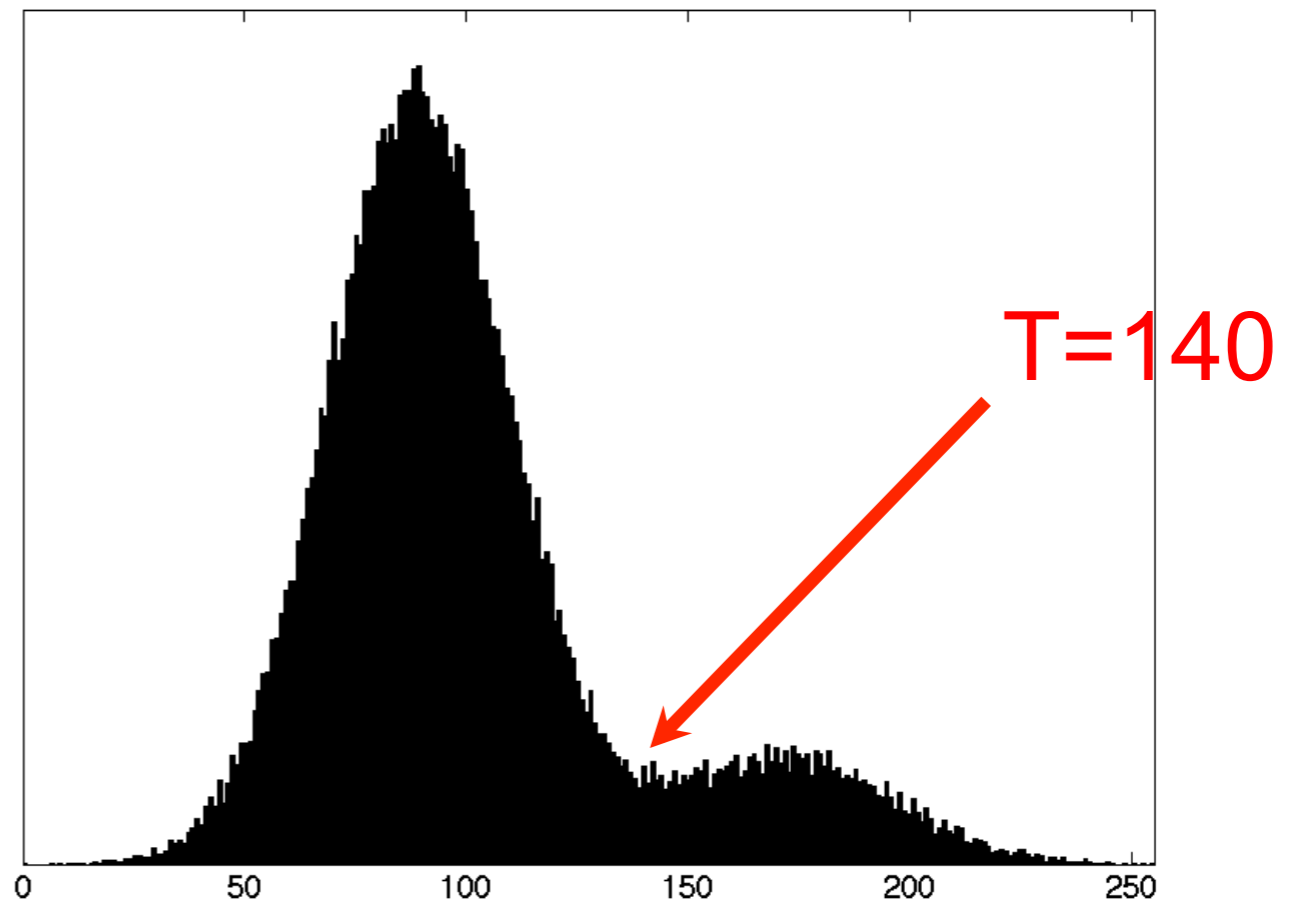
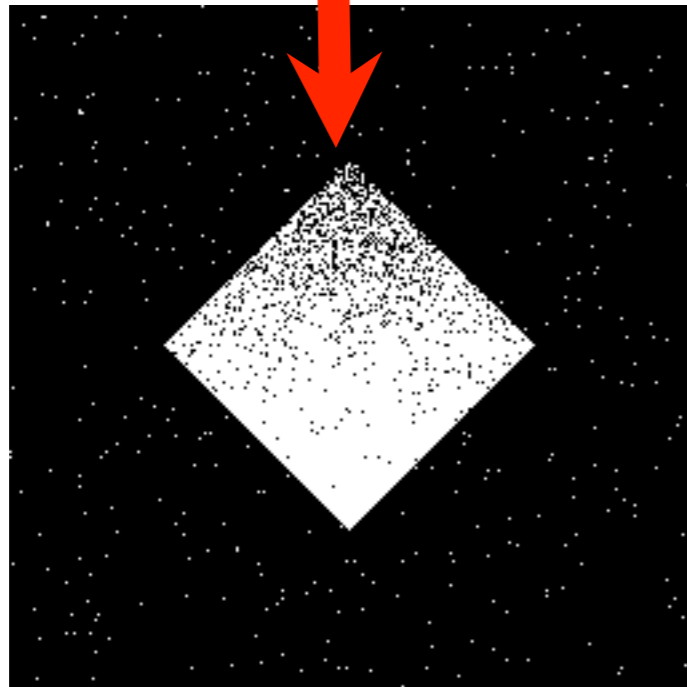
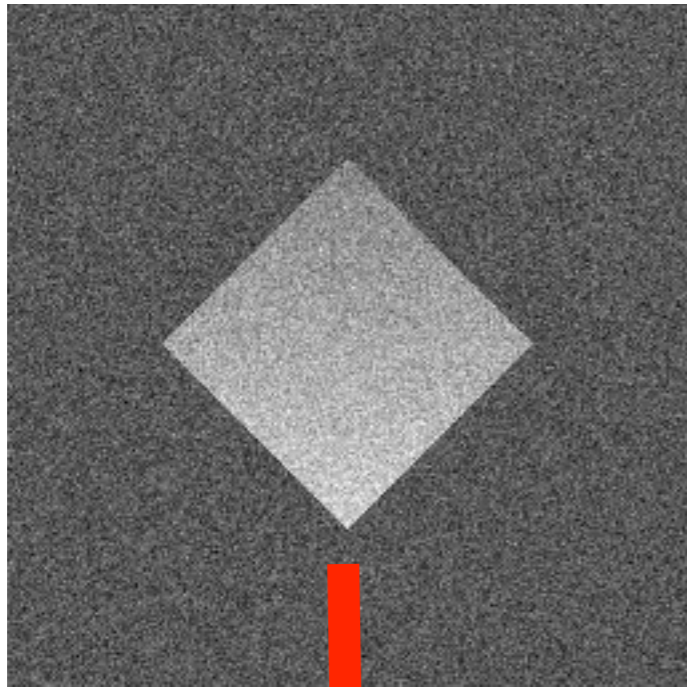
# Role of noise



# Low signal-to-noise ratio



# Low signal-to-noise ratio



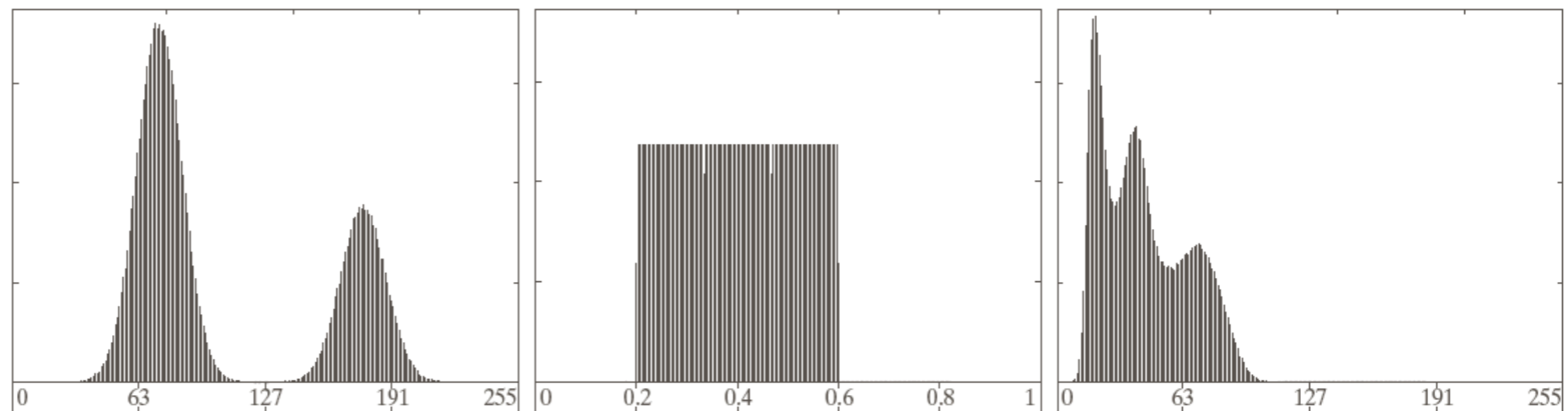
- How can we choose  $T$ ?
  - Trial and error
  - Use the histogram of  $f(x,y)$
  - Automatically
    - Otsu's method

# Effect of illumination on image histogram

Images



Histograms



$f$

$\times$

$g$

$=$

$h$

Original  
image

Illumination  
image

Final  
image

**45**

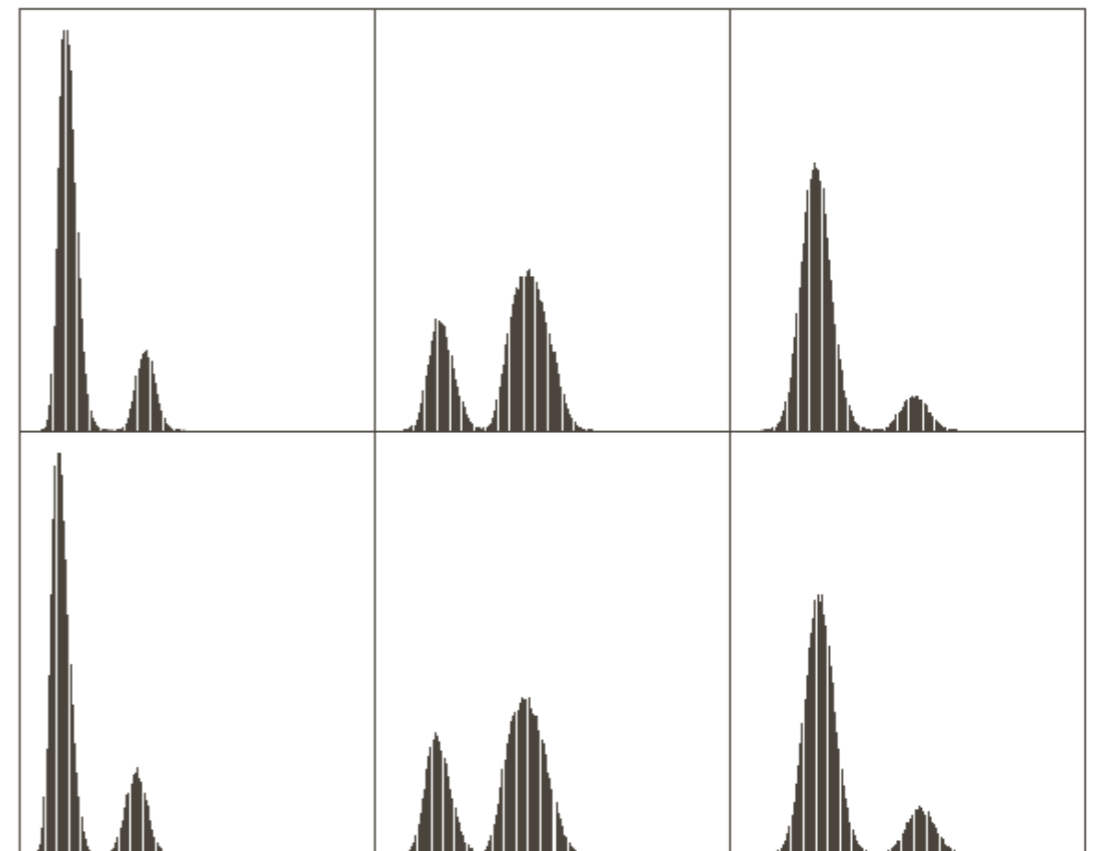
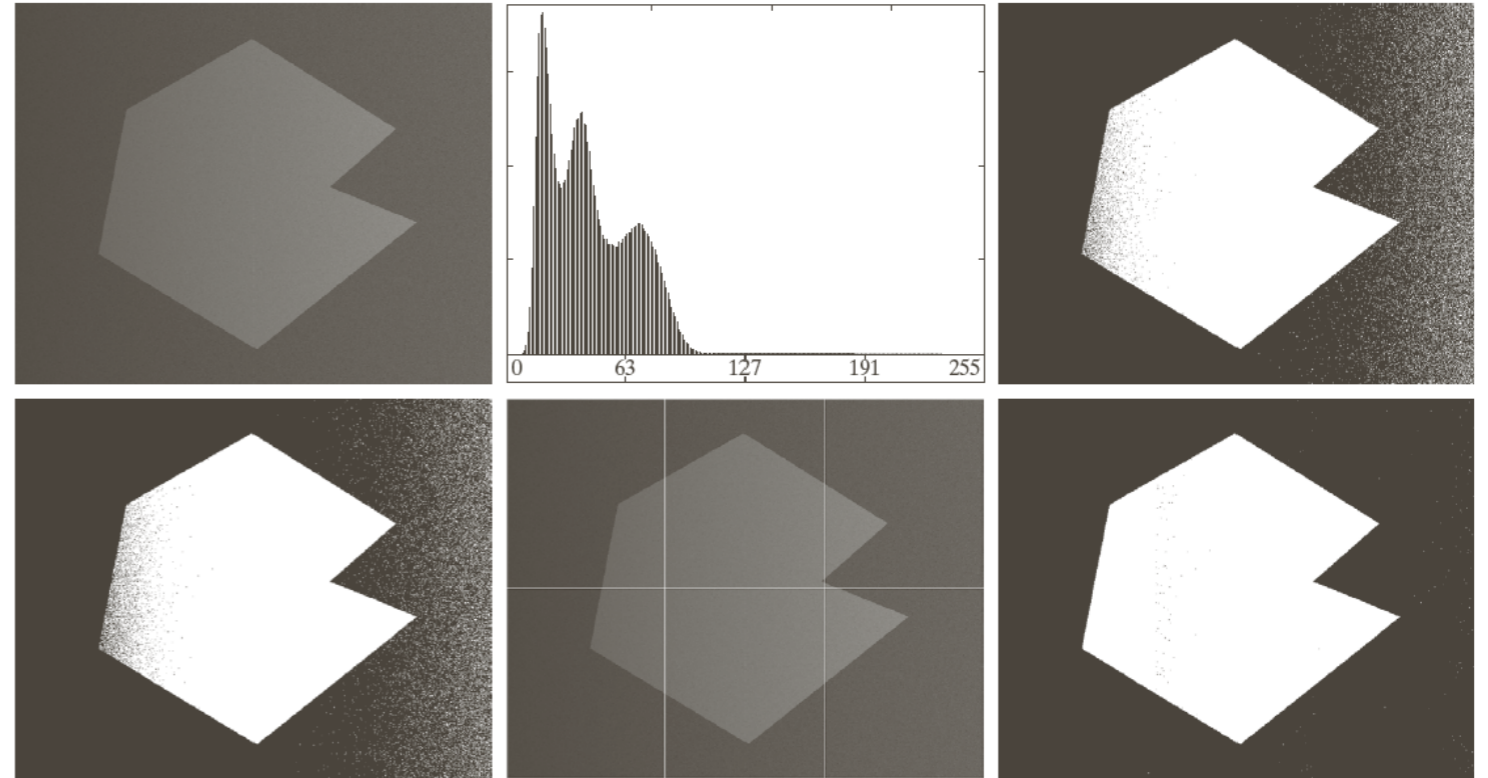
# Some applications

- Microscopy image showing bright cells on a dark background
  - Find number of cells
- Time-lapse microscopy images with lighting varying over time
  - Track number of cells over time



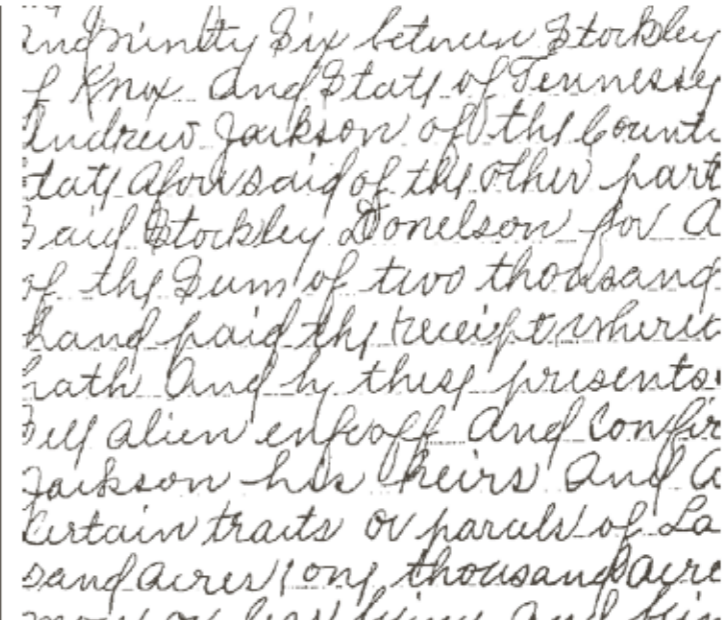
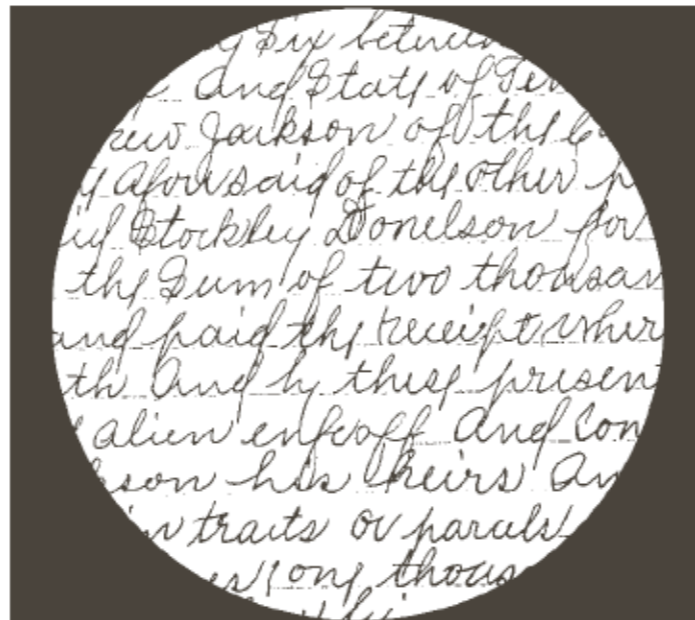
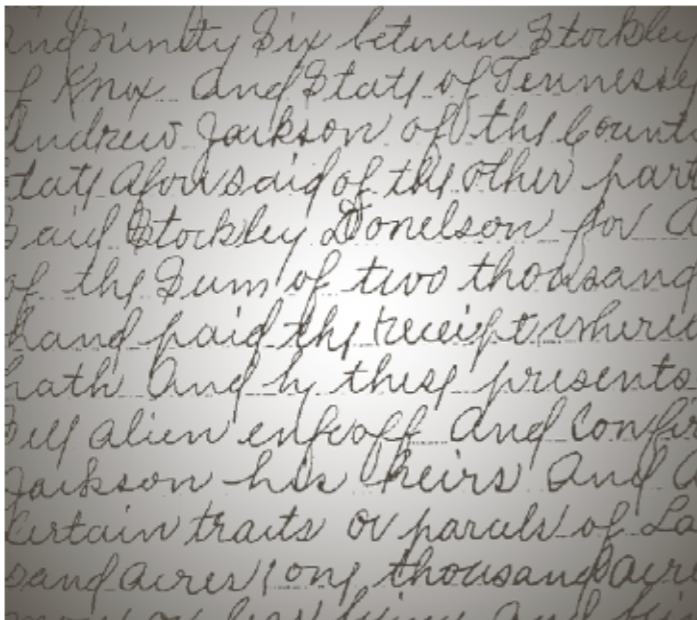
# Local thresholding

- One simple way
  - Subdivide image into blocks
  - Assumes every block has a portion of foreground and background



# Moving mean

- At every pixel  $(x,y)$  we can choose a threshold based on the mean  $m(x,y)$  of a local window
- This is very useful for adapting to changes in illumination
  - Can be problematic if the window at  $(x,y)$  contains only foreground or only background



# Moving mean - another example

- At every pixel  $(x,y)$  we can choose a threshold based on the mean  $m(x,y)$  of a local window
- This is very useful for adapting to changes in illumination
  - Can be problematic if the window at  $(x,y)$  contains only foreground or only background

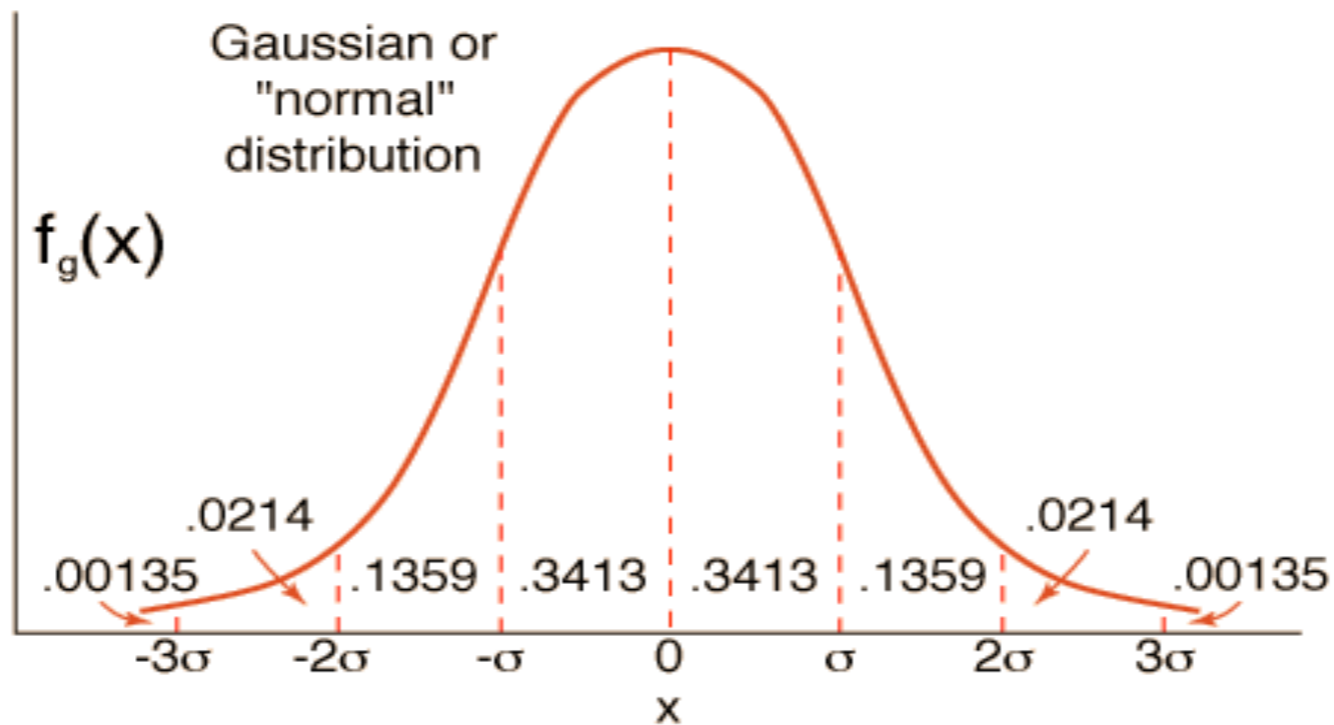


# Some Extra Things

- Gaussian/normal distribution
- Weighted means

# Gaussian Distribution

- “Normal” or “bell curve”
- Two parameters
  - $\mu$  = mean,  $\sigma$  = standard deviation



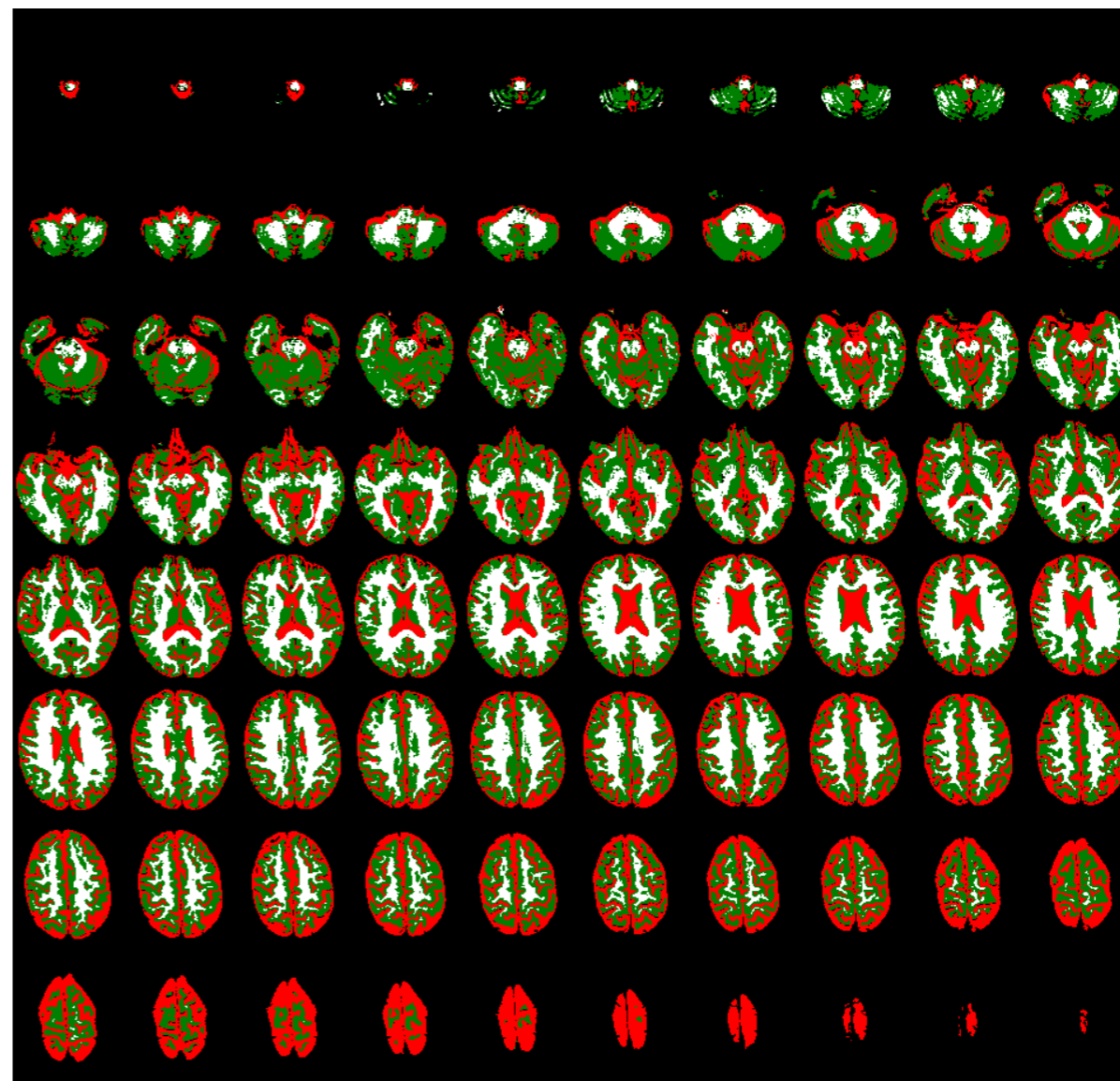
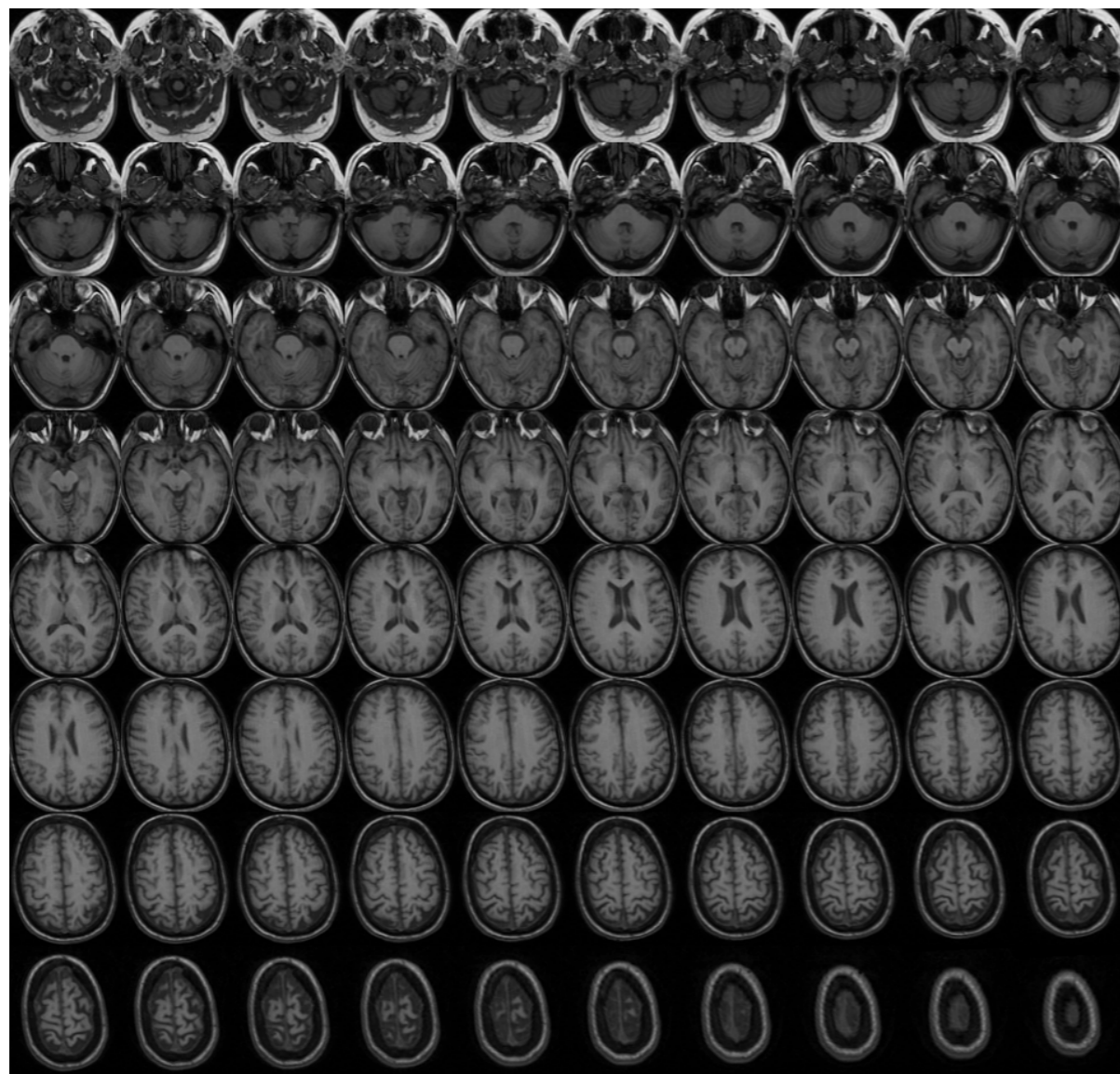
$$\frac{1}{\sigma\sqrt{2\pi}} \exp\left(-\frac{(x-\mu)^2}{2\sigma^2}\right)$$

# Gaussian Properties

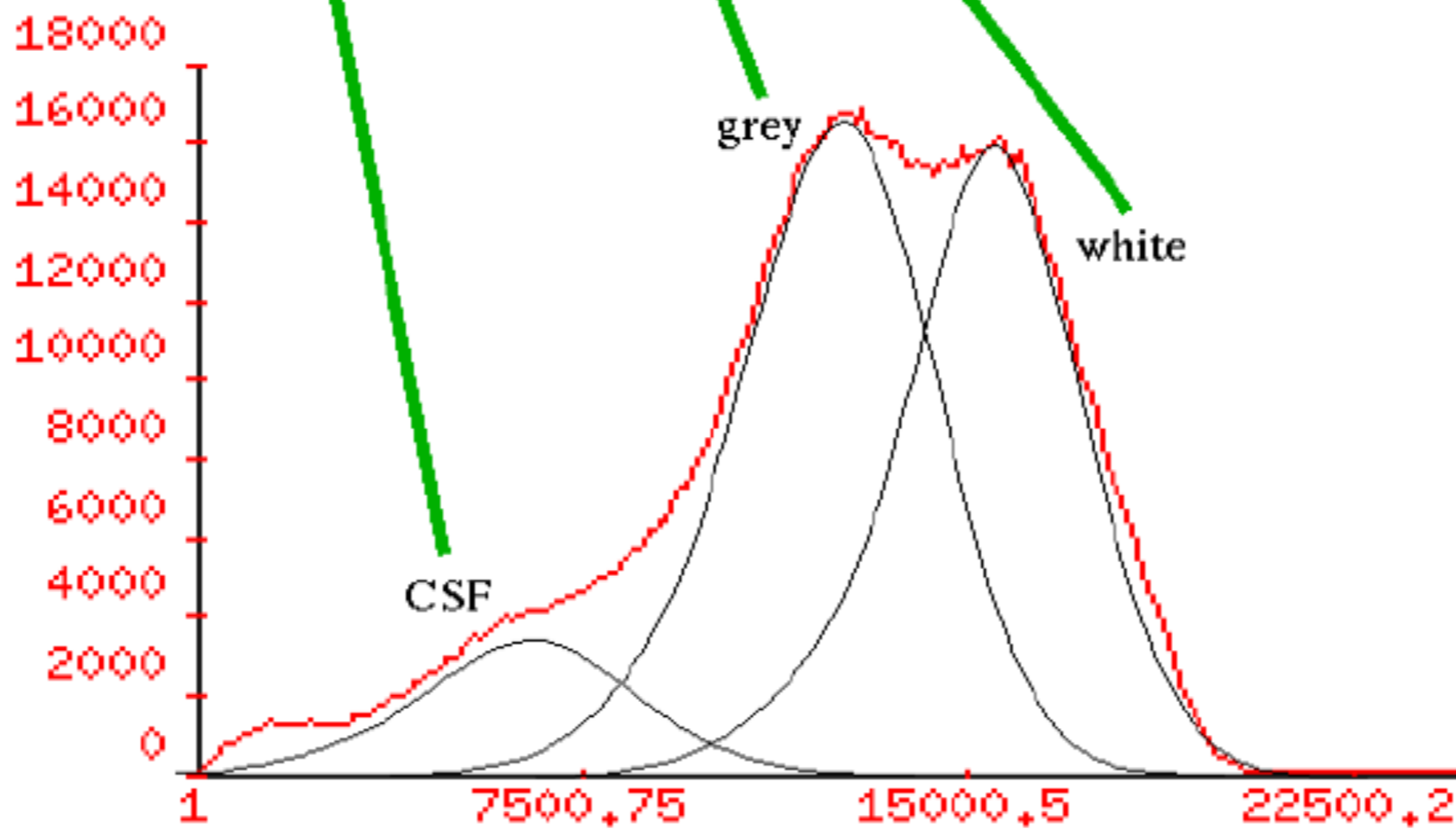
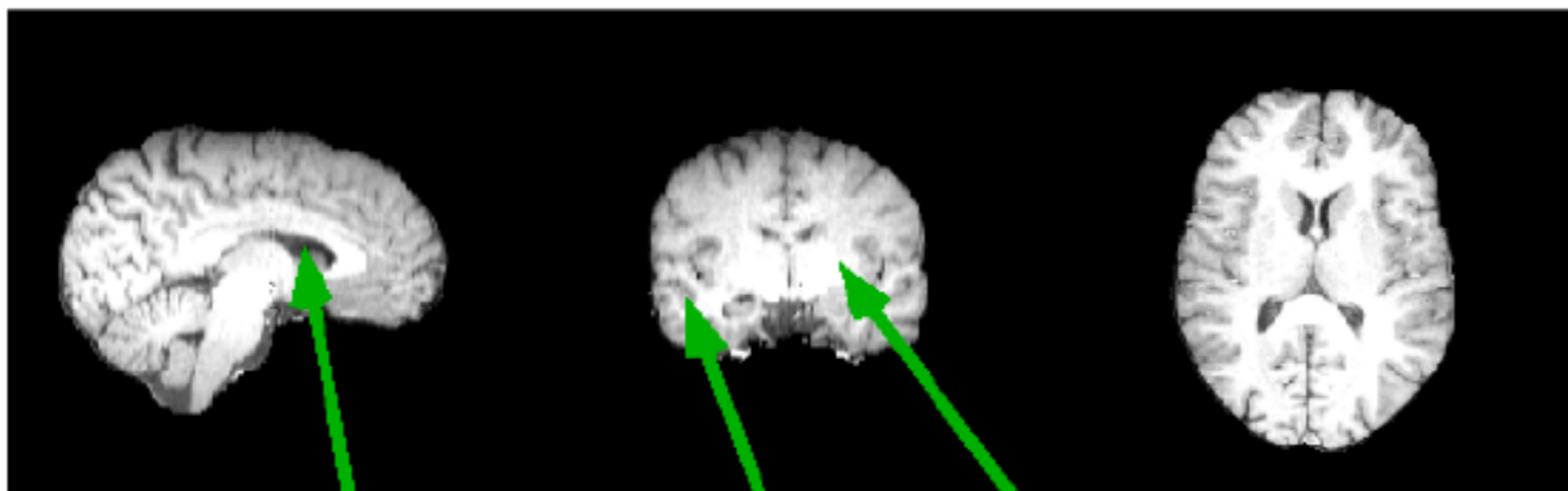
- Best fitting Gaussian to some data is gotten by mean and standard deviation of the samples
- Occurrence
  - Central limit theorem: mean of lots of independent & identically-distributed RVs
  - Nature (approximate)
    - Measurement error, physical characteristic, physical phenomenon
    - Diffusion of heat or chemicals

# MRI Brain Tissue Segmentation

- Segment gray matter, white matter, CSF and non-brain regions
  - Manual segmentation requires expertise and is very time consuming. It is also subjective.



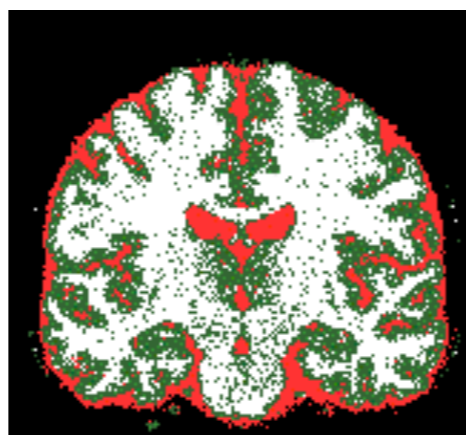
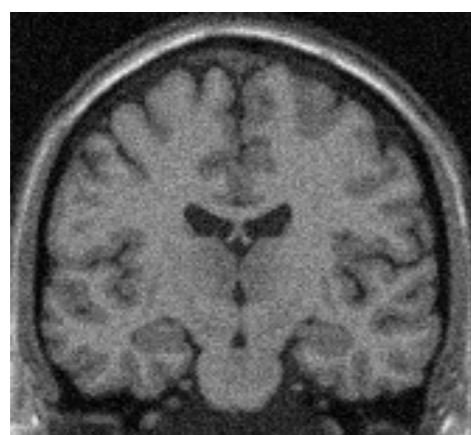
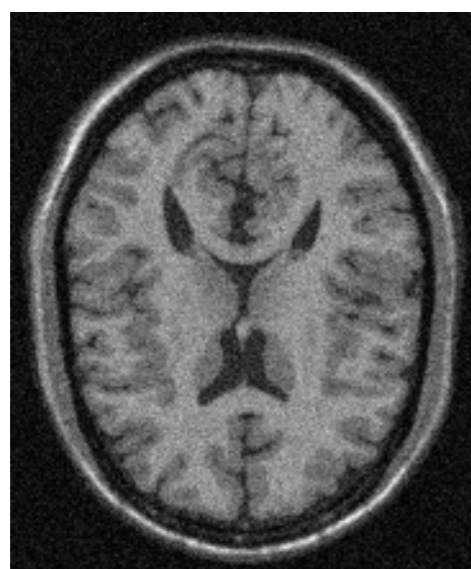
# MRI Brain Example





# Effect of noise

- Overlap in conditional probabilities of different classes.

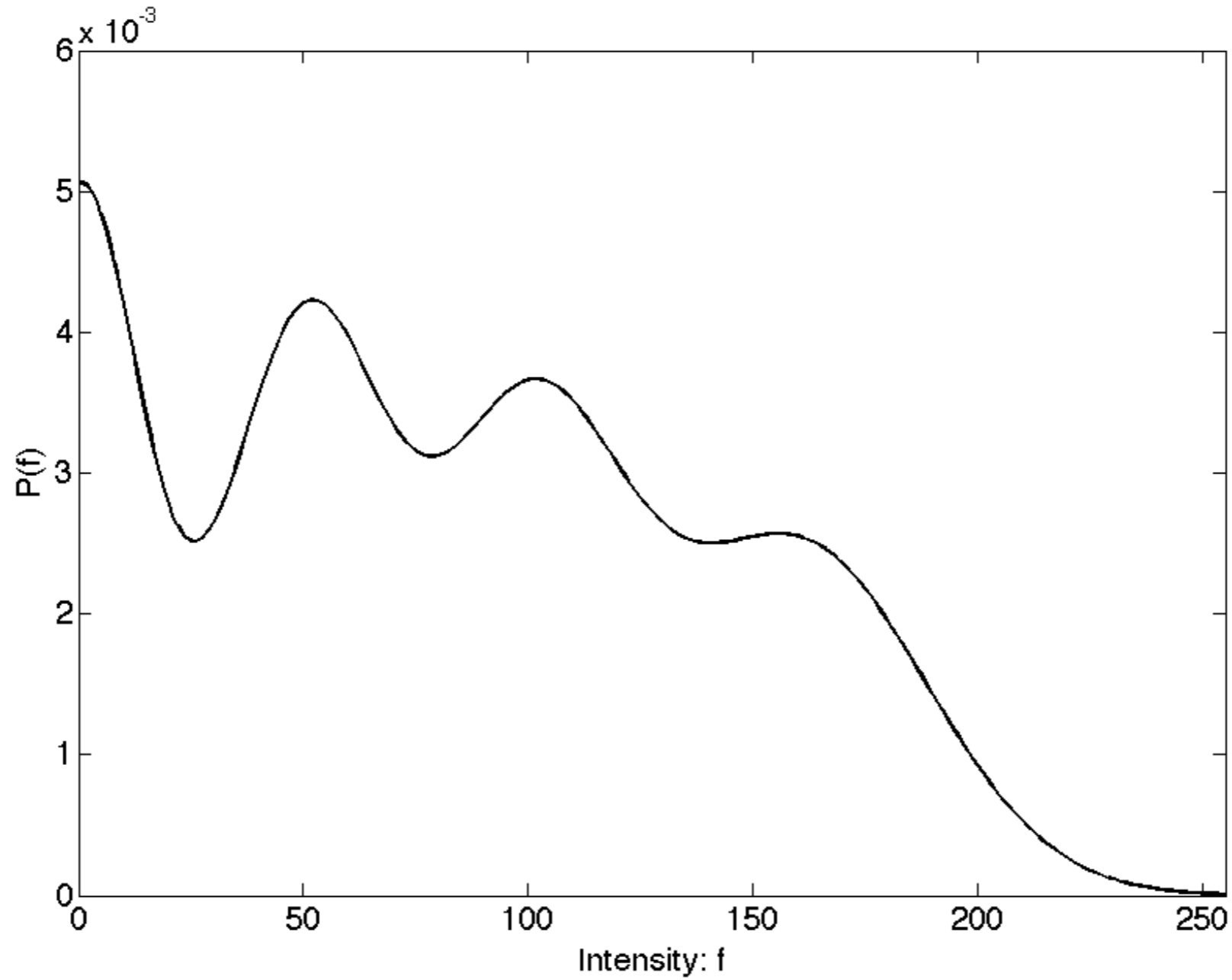


Image

Thresholding only

With spatial filtering

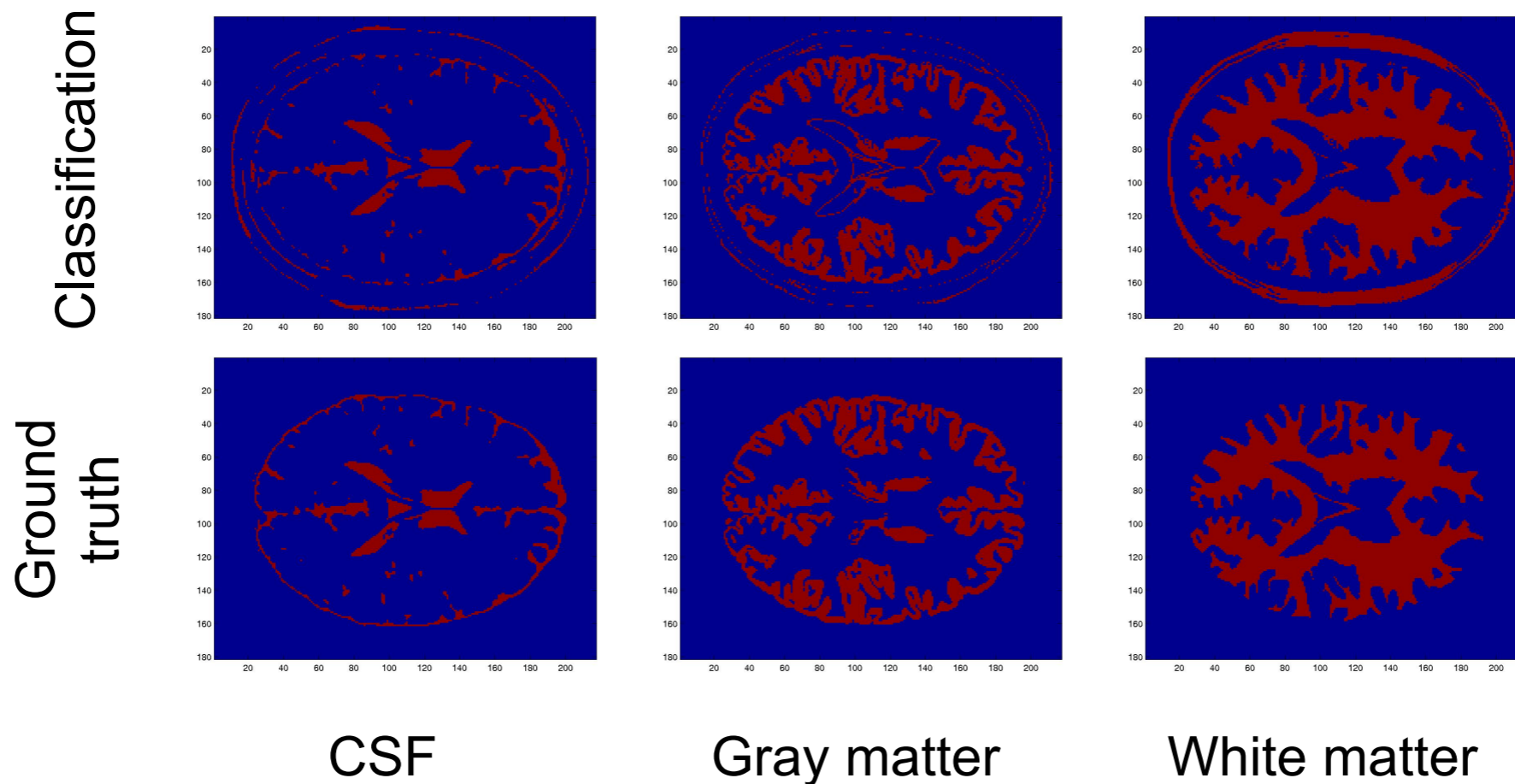
# Observed image histogram



$$P(r_k) = \sum_{l \in \{csf, gm, wm, background\}} P(r_k | l) P(l)$$

# Non-brain tissues

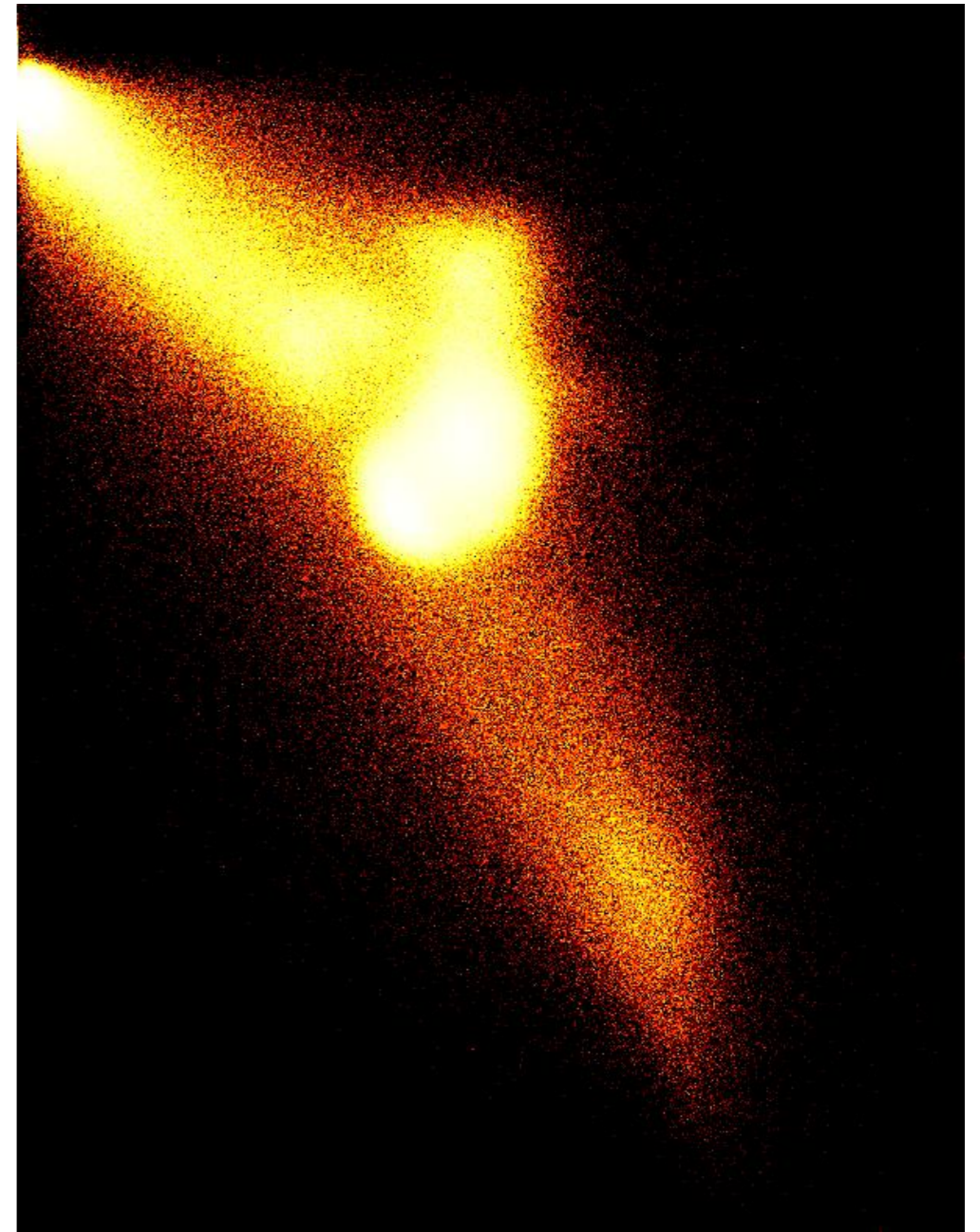
Large amount of overlap brain vs. non-brain



When conditional probabilities overlap this significantly, simple thresholding techniques fail. There are several possible solutions...

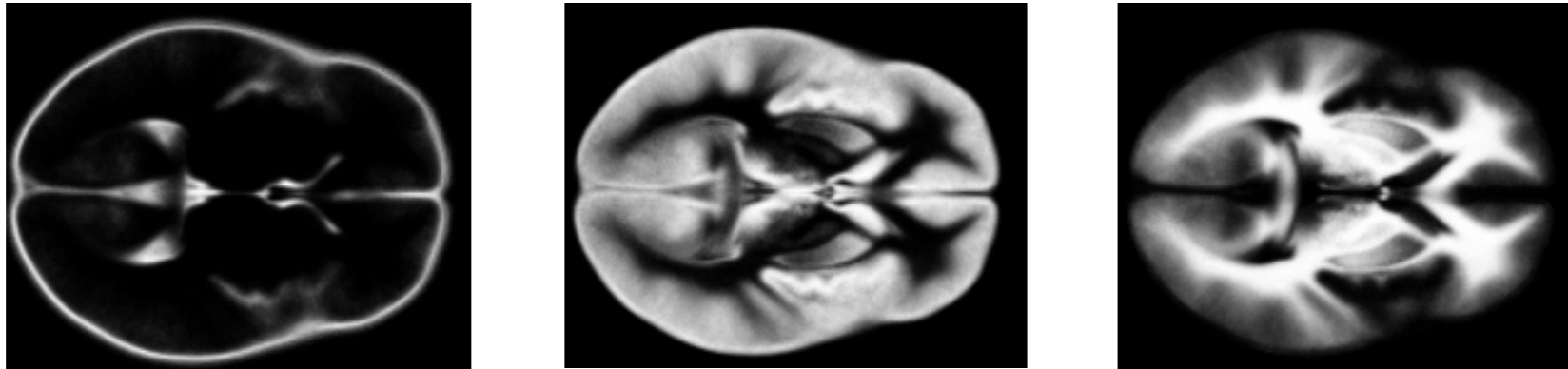
# Multidimensional histograms

- Sometimes we have multiple images of the same object
  - Different channels in multispectral data
  - Magnetic resonance images with different pulse sequences
- Each channel provides new information
- If we have two channels, we can create a 2D histogram



A 2D histogram of a brain image from 2 MRI channels

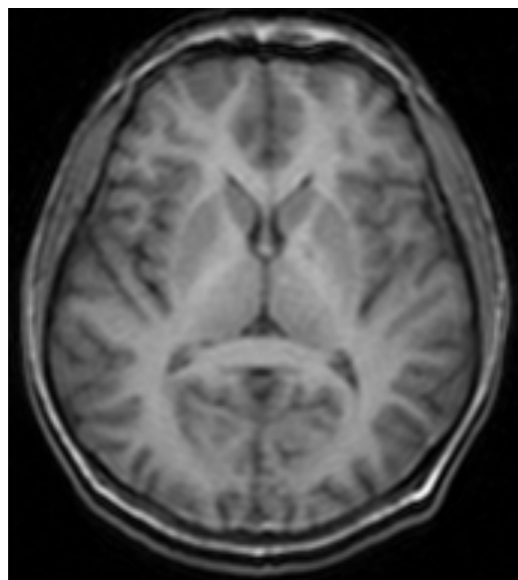
# Non-brain tissues



- One solution is to use atlases to guide the segmentation in addition to thresholding
- 452 subjects scanned
  - Manually segmented
  - Images aligned into common coordinate system
  - Atlas: tissue classification probability images
    - Gives a prior probability for all tissue classes at each voxel
- When we have a new MRI to segment
  - Register this atlas onto new subject's image
  - Use atlas to clarify ambiguities

# Effect of non-homogeneous intensity

- Magnetic resonance images typically have a multiplicative bias field
  - This is similar to variable illumination
  - Unfortunately, the bias field can depend on the object being imaged!
  - Fortunately, it can still be estimated



Image



Thresholding only



With bias field correction